Acute Toxicity Assessment of Sodium Arsenite in Zebrafish (*Danio rerio*) Under Ideal Physico-Chemical Conditions

Divya Prasad¹, Sukriti Suman², Vikash Kumar³, Kundan Patel⁴ and Gyanendra B. Chand⁵*

^{1, 2, 4, 5}Aquatic Toxicology Laboratory, P.G. Department of Zoology, Patna University, Patna – 800005, Bihar, India ³ Department of Biochemistry, Rajendra Memorial Research Institute of Medical Sciences, Patna – 800007, Bihar, India

S*Corresponding Authors, Corresponder D. Chand

^{5*}Corresponding Author: Gyanendra B. Chand

Abstract:- Arsenic is ubiquitously prevalent metalloid contaminant released in the aquatic environment as a consequence of geogenic and anthropogenic activities. After West Bengal, Bihar has emerged as a new hotspot for arsenic toxicity in India. Presence of arsenic in groundwater and soil in various forms has been a matter of concern for the scientific community globally. The present study aims to assess the median lethal concentration (LC₅₀) for sodium arsenite (NaAsO₂) in Danio rerio (zebrafish). Healthy fish were procured from Ashish Aquarium Traders, Howrah, India, acclimated in the laboratory for 15 days in ideal laboratory condition and fed *ad libitum*. Following a range finding test for arsenic, a set of doses were administered to the fish for 24h, 48h, 72h and 96h. The LC₅₀ for each assessment period was separately determined using probit regression analysis and later confirmed by pilot test. The24h, 48h, 72h and 96h LC50were determined as 45mg/L, 44mg/L, 41mg/L and 39mg/L respectively. Since there is great similarity in genome of zebrafish and humans, the finding of the present acute toxicity study may be interpreted for the humans.

Keywords:- Acute Toxicity, Anthropogenic, Danio Rerio, Geogenic, LC₅₀, Physic-Chemical, Probit Regression, Sodium Arsenite.

I. INTRODUCTION

Aquatic ecosystem has been subjected to immense anthropogenic load in terms of non-judicious applications of agrochemicals including various synthetic heavy metals[1][2][3][4]. Heavy metals are those that have a relatively high density compared to water $(>5g/cc^3)$ and their atomic weight is 63.5-200.5 g/mol[5]. Heavy metal contamination of the environment results from both natural sources and human activities. They are not completely metabolised by the living system and as such they accumulate in body tissues and disturb normal physiology. In fishes they take their entry via general body surface, gills or digestive tract and induces various physiological, biochemical and oxidative perturbations[1][6][7].

Weathering of rock, leaching, run off, volcanic and biological activities include the natural resources of fresh water contamination[8][9]. Major anthropogenic sources of

groundwater arsenic contamination are extensive mining and geothermal activities, fossil fuel combustion, use of metallic arsenite for strengthening alloys as well as in the processing of glass pigments, textiles, metal adhesive wood preservatives, ammunition and other synthetic arsenical compounds[10][11]. More than 230 million people worldwide are suffering due to arsenic contaminated consumption of water including 180 million people from Asia alone. In India, around 50 million people living in 20 states have been reported to be affected by arsenic contaminated groundwater consumption[12]. The soil and groundwater concentration of arsenic in some of these states adjacent to Ganga-Brahmaputra basin have been found to be above the limit specified far bv WHO *i.e.*,10ppb[13][14][15]. It is known to cause gall bladder, urinary bladder, skin and liver cancer and various other physiological abnormalities. It is known to disrupt physiological processes via induction of oxidative stress. It creates imbalance between RONS (reactive oxygen and nitrogen species) generation and annihilation. Arsenic induced oxidative stress is reported to damage cell membrane and DNA while also affecting major cell organelles including Golgi complex and ER[7][16][17][18].

In nature, arsenic is present as both arsenite (3+) and arsenate (5+). Arsenite is reported to be comparatively more toxic form[12]. Since fishes are best sentinels to study the pollution status of an aquatic body, they have been widely used to assess water quality of aquatic ecosystem[19][20]. Following heavy metal toxicity, the growth rate, reproductive efficacy and other metabolic functions of the fish are hindered, ultimately causing mortality in fishes[21].

Zebrafish is a tropical freshwater fish in the minnow family. Since past few decades, it has become a preferred model organism for human disease research as 70% of the protein-coding human genes are related to genes found in zebrafish[22]. Several research has revealed that zebrafish undergo similar changes in its enzymatic profile as humans when exposed to heavy metals[23][16][24]. The present study has been carried out to assess the acute toxicity of sodium arsenite in zebrafish in ideal laboratory conditions. It is also focussed on the study of behavioural changes and corresponding stress responses of the fish during acute toxicity test.

ISSN No:-2456-2165

II. MATERIAL AND METHOD

A. Experimental animal:

Danio rerio with an average weight of $0.409 \pm 0.02g$ and length 3.5 ± 0.05 cm were purchased from Ashish Aquarium Traders, Howrah, India, in June 2022. Fish were given a bath of 0.1% KMnO₄ solution, segregated as per size in 20L aquaria and acclimated for 15 days in the laboratory. They were fed daily *ad libitum*.

B. Experimental Design:

A total of 240 fish were used in the experiment. The fish were divided into 8 groups of 10 fish each in 20L aquarium capacity- 7 experimental groups and 1 control group. The test was performed in triplicate.

The physico-chemistry of water *i.e.*, temperature, pH, dissolved oxygen were analysed as per the standard protocol of APHA (2017).

C. Preparation of stock solution and dose administration:

Sodium arsenite (NaAsO₂, 98.9% pure) was purchased from Loba Chemie Pvt. Ltd. Stock solution of 20,000 mg/L was prepared by dissolving NaAsO₂ in distilled water. The test solution used in 7 different groups ranged from 20mg/l to 50mg/l. The feeding of the fish was stopped during the test. The test solution was renewed every day in order to maintain the desired concentration in each group. The fish were examined closely for any behavioural or morphological changes. The dead fish were removed immediately to prevent the contamination of water.

D. Statistical Analysis:

The percentage mortality at 24, 48, 72 and 96 hours were recorded. The corresponding LC_{50} values and its confidence limits (95% CL_8) were calculated by Finney's probit regression method and confirmed by pilot test. To determine the relationship between logarithm of concentration and the corresponding percentage mortalities, the regression equation was obtained using MS Excel Office 2021 software. The antilog of concentration gave the value of LC_{50} . All tests were performed in triplicate.

III. RESULTS AND DISCUSSION

To study the acute toxicity of sodium arsenite in *Danio rerio*, doses of 20, 25, 30, 35, 40, 45 and 50 mg/L of the test chemical were administered to the fish of different aquaria. The percentage mortalities were recorded at the end of 24 h, 48 h, 72 h and 96 h (table1 and figure 1).

Table 1. Percentage death of fish exposed to different concentrations of sodium arsenite for different durations.

Conc.	Log conc.	%	Probit	%	Probit	% mortality	Probit	%	Probit
(mg/L)		mortality	kill	mortality	kill	In 72 h	kill	mortality	kill
		in 24 h		In 48 h				In 96 h	
20	1.30103	0	0.00	0	0.00	0	0.00	0	0.00
25	1.39794	10	3.72	0	3.72	0	3.72	0	3.72
30	1.47712	20	4.16	0	4.16	0	4.16	10	4.48
35	1.54406	20	4.16	0	4.16	10	4.48	10	4.75
40	1.60206	30	4.48	0	4.48	10	4.75	10	5.00
45	1.65321	20	4.16	0	4.16	20	4.75	10	5.00
50	1.69897	60	5.25	10	5.52	10	5.84	0	5.84

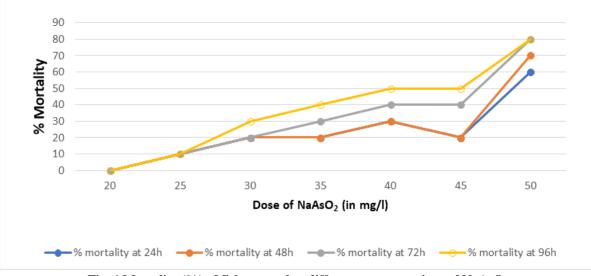
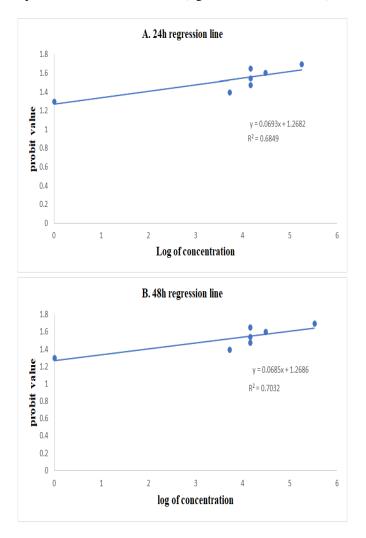


Fig. 1 Mortality (%) of fish exposed to different concentrations of NaAsO2

S.No	Exposure period	LC ₅₀ (mg/L)		
1.	24 hours	45.289		
2.	48 hours	44.400		
3.	72 hours	41.209		
4.	96 hours	39.719		

It is evident that the mortality is positively correlated with dose. There was no mortality in the control group. There is marked difference in LC_{50} values at each successive 24 hour. The percentage mortality was converted into probit units and plotted against the logarithm of concentration of sodium arsenite to obtain a regression line and regression equation for each time interval (figure 2A, 2B, 2C and 2D).



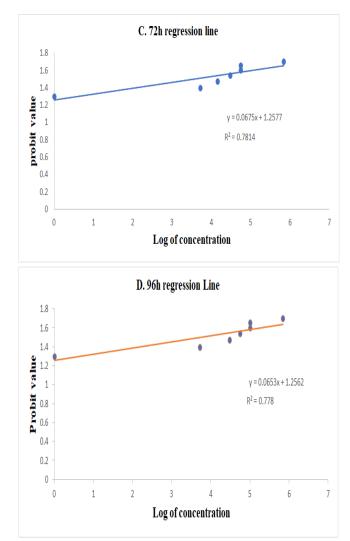


Fig 2. Regression line between probit kill and log concentration for different time intervals [A-24h, B-48h, C-72h, D-96h] in NaAsO₂ treated zebrafish.

Zebrafish, belonging to Cyprinidae family, is a popular freshwater fish native to South Asia. In recent years it has become one of the most preferred model organisms due to various reasons[24]. The present study revealed the 24h, 48h, 72h and 96h LC₅₀ value of sodium arsenite to the fish as 45 mg/L, 44 mg/L, 41 mg/L and 39 mg/L respectively (table 2).

The findings of the present investigation show doseand duration-dependent mortality in zebrafish. At 48 hours there is slight decline in mortality from 30% to 20% as the dose increases from 40mg/L to 45mg/L, suggesting an increase in oxidative stress resistance mechanism in fish after a certain threshold dose. The anti-oxidants might have attained a peak following 48-hour exposure to 40mg/L of

ISSN No:-2456-2165

sodium arsenite, which was the probable cause for decline in mortality till an exposure to 45mg/L. The fish failed to counter oxidative stress generated by further higher concentration of the toxicant exposure and therefore there has been no decline in mortality following 48 hours.

The acceptable limit of total arsenic concentration is 340 μ g/L for acute exposure and 150 μ g/L for chronic exposure in freshwater as per USEPA aquatic life criteria. However, according to NAS (1977), a concentration of 1.5-3.8 mg/L of arsenite has been considered safe for the fish. The concentration of 13.6 mg/L of As (III), used for effective weed control has been considered to be harmful to several fish species[25][26].

Acute toxicity is the discernible adverse effect induced by a substance in an organism within a short time of exposure[21][27]. It is the effect that can damage organs due to exposure of chemical/ toxicant in a short time[28].

IV. CONCLUSION

The findings of the present study provide an insight of acute toxicity range of sodium arsenite to the fresh water fish *Danio rerio*. The LC₅₀ value determined in the present acute toxicity assessment is much higher than their environmental relevance, still it contributes a lot in determination of dose and duration of AS(III) to the freshwater fish while studying the molecular mechanism related to oxidative and genotoxic potentials of the toxicant to the fish. Acute toxicity test is highly appreciated to predict the safe concentration of a particular toxicant in the environment.

Source of Funding

This research did not benefit from grant from any nonprofit, public or commercial funding agency.

ACKNOWLEDGEMENT

The authors are thankful to Head, Department of Zoology, Patna University, Patna for providing infrastructural facilities.

Conflict of Interest

The authors declare no conflict of interest.

REFERENCES

- M. A. Mandour, S. A. Al-Shami, and Y. A. Hssein, "Effect of feeding graded levels of urea on growing New Zealand white rabbit performance," *Glob. Vet*, vol. 9, no. 6, pp. 761–768, 2012.
- [2]. A. Ghaffar *et al.*, "Clinico-hematological disparities induced by triazophos (organophosphate) in Japanese quail," *Pak. Vet. J.*, 2014.
- [3]. J. Mashkoor, A. Khan, M. Z. Khan, M. K. Saleemi, and F. Mahmood, "Arsenic induced clinico-hematopathological alterations in broilers and its attenuation by vitamin E and selenium," *Pakistan J. Agric. Sci.*, 2013.

- [4]. M. Witeska, P. Sarnowski, K. Ługowska, and E. Kowal, "The effects of cadmium and copper on embryonic and larval development of ide Leuciscus idus L," *Fish Physiol. Biochem.*, 2014.
- [5]. M. Koller and H. M. Saleh, "Introductory Chapter: Introducing Heavy Metals," in *Heavy Metals*, 2018.
- [6]. M. A. Abdl-rahman, S. Y. Saleh, A. Zaki Amal, and S. A. El-Hamid Safaa, "Growth Performance, Cecal Fermentation and Blood Biochemistry of Rabbits Fed Diet Supplemented with Urea Bentonite Combination," J. Agric. Sci., 2011.
- [7]. G. B. Chand and S. Kumar, "Arsenic Induced Oxidative Perturbations in Freshwater Air Breathing Fish Clarias batrachus: In Vivo Study," *Jordan J. Biol. Sci.*, 2020.
- [8]. M. Azizur Rahman and H. Hasegawa, "Arsenic in freshwater systems: Influence of eutrophication on occurrence, distribution, speciation, and bioaccumulation," *Appl. Geochemistry*, 2012.
- [9]. W. Zhang, Y. Liu, M. Ge, C. Yao, J. Xue, and Z. Zhang, "Resveratrol reduces oxidative stress and improves arsenic efflux in rats exposed to arsenic trioxide," *Pak. Vet. J.*, 2014.
- [10]. P. L. Smedley and D. G. Kinniburgh, "Arsenic in groundwater and the environment," in *Essentials of Medical Geology: Revised Edition*, 2013.
- [11]. S. A. A.-W. Al-Sulaiman, "Chemical Safety of Drinking-Water: Assessing Priorities for Risk Management," *Int. J. Environ. Stud.*, vol. 69, no. 6, pp. 1001–1001, 2012.
- [12]. E. Shaji, M. Santosh, K. V. Sarath, P. Prakash, V. Deepchand, and B. V. Divya, "Arsenic contamination of groundwater: A global synopsis with focus on the Indian Peninsula," *Geosci. Front.*, 2021.
- [13]. R. Nickson *et al.*, "Current knowledge on the distribution of arsenic in groundwater in five states of India," *J. Environ. Sci. Heal. Part A Toxic/Hazardous Subst. Environ. Eng.*, 2007.
- [14]. R. Singh, S. Singh, P. Parihar, V. P. Singh, and S. M. Prasad, "Arsenic contamination, consequences and remediation techniques: A review," *Ecotoxicology and Environmental Safety*. 2015.
- [15]. D. Ghosh and J. Routh, "DISSOLVED ORGANIC CARBON IMPACT MICROBIAL ARSENIC CYCLING IN AQUIFERS OF WEST BENGAL (INDIA) DISSOLVED ORGANIC CARBON IMPACT MICROBIAL ARSENIC CYCLING IN AQUIFERS OF WEST BENGAL (INDIA)," no. October, pp. 10–11, 2014.
- [16]. P. Delaney, A. Ramdas Nair, C. Palmer, N. Khan, and K. C. Sadler, "Arsenic induced redox imbalance triggers the unfolded protein response in the liver of zebrafish," *Toxicol. Appl. Pharmacol.*, 2020.
- [17]. J. H. Huang and G. Ilgen, "Blank values, adsorption, pre-concentration, and sample preservation for arsenic speciation of environmental water samples," *Anal. Chim. Acta*, 2004.
- [18]. A. Weidinger and A. V. Kozlov, "Biological activities of reactive oxygen and nitrogen species: Oxidative stress versus signal transduction," *Biomolecules*. 2015.

- [19]. I. Naigaga *et al.*, "Fish as bioindicators in aquatic environmental pollution assessment: A case study in Lake Victoria wetlands, Uganda," *Phys. Chem. Earth*, 2011.
- [20]. L. S. Balistrieri, S. E. Box, A. A. Bookstrom, R. L. Hooper, and J. B. Mahoney, "Impacts of historical mining in the Coeur d' Alene River Basin," *Pathways Met. Transf. from Miner. Sources to Bioreceptors A Synth. Miner. Resour. Program'sPast Environ. Stud. West. United States Futur. Res. Dir.*, pp. 1–34, 2002.
- [21]. A. Ullah *et al.*, "Determination of 96-hr LC 50 value of cadmium for a fish, Labeo rohita," ~ 380 ~ J. Entomol. Zool. Stud., 2016.
- [22]. P. Mondal *et al.*, "Combined effect of arsenic and fluoride at environmentally relevant concentrations in zebrafish (Danio rerio) brain: Alterations in stress marker and apoptotic gene expression," *Chemosphere*, 2021.
- [23]. P. Carlson, D. M. Smalley, and R. J. Van Beneden, "Proteomic analysis of arsenic-exposed zebrafish (danio rerio) identifies altered expression in proteins involved in fibrosis and lipid uptake in a genderspecific manner," *Toxicol. Sci.*, 2013.
- [24]. J. Hallauer, X. Geng, H. C. Yang, J. Shen, K. J. Tsai, and Z. Liu, "The Effect of Chronic Arsenic Exposure in Zebrafish," *Zebrafish*, 2016.
- [25]. "Effects of arsenic in the Canadian environment," 1978.
- [26]. E. M. B. Sorensen, R. Ramirez-Mitchell, A. Pradzynski, T. L. Bayer, and L. L. Wenz, "Stereological analyses of hepatocyte changes parallel arsenic accumulation in the livers of green sunfish," J. Environ. Pathol. Toxicol. Oncol., 1985.
- [27]. Kundan P., Mohan, K., & G B Chand, "Acute toxicity assessment of sodium fluoride to freshwater air breathing fish Channa punctatus: in vivo study," *Anim. Sci.*, vol. 17, no. 2, pp. 89–92, 2022.
- [28]. I. M. Widiastuti, A. M. S. Hertika, M. Musa, and D. Arfiati, "Acute toxicity test and LC50 value of mercury on tubifex tubifex," in *Journal of Physics: Conference Series*, 2019.