

Review: Role of NONO Gene from DBHS Complex and its Interaction with PIN1 in Cancer

Sayed Tariq Pachakhan^{*1,2}, Hasamudin Sayedi³, Samira Hamidi⁴

¹ Research Directorate and Lecturer, Spinghar Institute of Higher Education (Kabul campus) Afghanistan

² Laboratory Head of Afghan Japan communicable disease hospital.

³ Dean of Medical Laboratory Sciences Spinghar Institute of Higher Education (Kabul campus) Afghanistan

⁴ Assistant Professor Department of Molecular biology, Kabul University of Medical Sciences

Abstract:- Paraspeckle are sub-nuclear bodies found in the interchromatin space of mammalian cells, made from the interaction of non-coding RNA and DBHS (Drosophila behavior human splicing) complex. DBHS consists of p54NRB/NONO, PSPC1, and PSF/SFPQ proteins. Each protein of the DBHS complex contains RRM1/2, NOPS, and the Coiled-coil domain. Amongst the DBHS complex, NONO protein is involved in breast, prostate, and malignant melanoma progress. NONO gene is engaged in various biological processes, including RNA splicing and editing, DNA unwinding and repairing, and gene transcription. NONO can interact with SFPQ and PSPC1 in the DBHS complex as heterodimers and homodimers. In addition, NONO can also interact with Sterol regulatory-element binding proteins (SREBPs) and stabilize them. (SREBPs) activate genes encoding key enzymes responsible for the biosynthesis of fatty acid and cholesterol in breast cancer. PIN1 and SUPT5H are oncogenic proteins; the function of PIN1 is defined by its two domains: the WW domain and the PPIase domain. NONO and SUPT5H interact with the WW domain of the PIN1 protein, which induces the PPIase domain and promotes cancer. PIN1 stabilizes the NONO, and SUPT5H, contributing to breast cancer cell proliferation, migration, invasion, cell cycle, and apoptosis. Silencing NONO, PIN1, and SUPT5H can suppress tumor growth and be a good target for therapeutic purposes.

Keywords:- NONO gene, DBHS Complex, PIN1, Cancer.

I. INTRODUCTION

We focused on the functional interaction of Non-POU Domain-Containing Octamer-Binding (NONO) and DBHS complex protein and their correlation with cancer and interaction with PIN1. Paraspeckles are nucleosome bodies consisting of one non-coding RNA and a combination of DBHS proteins. DBHS is a family of three different proteins involved in distinct functions, NONO, PSPC1, and SFPQ [1]. DBHS proteins as multifunctional nuclear proteins implicated in subnuclear body formation, co-activation, transcription initiation [2][3] corepression, constitutive and alternative splicing [4][5][6][7] transcriptional termination [8] DNA repair [9], contributing to circadian rhythm regulation [10][11], tumor suppression [12] and Drosophila behavior [13]. From the DBHS complex, splicing factor proline and glutamine-rich (SFPQ) and NONO are highly plentiful and expressed in various cell lines and tissues [9]. DBHS

proteins are essential for forming subnuclear paraspeckles [14]. Crystallography results show that PSPC1 residues (61-32) and NONO residues (53-312) include a partial DBHS domain; it contains two RNA-recognition motifs (RRM) domains, the NONA/paraspeckle domain (NOPS) domain and part of the coiled-coil domain [15]. Each unit of DBHS protein has multiple binding domains which bind to a specific target. For NONO protein, these domains consist of n-terminal G-rich amino acid followed by RRM1, RRM2, NOPS, COILED-COIL domain and finally ending by GP-rich amino acid [16], [17]. The same lineup of domains is repeated for PSPC1 and SFPQ proteins; however, they are different at the start and end points [15]. PSPC1 has often been used as a paraspeckles marker [1]. In the DBHS complex, PSPC1 is much less abundant than SFPQ, thus, knocking down *pspc1* does not affect the paraspeckle complex within the HeLa cell. For the integrity and function of paraspeckles, the interaction between NONO and PSPC1 is crucial [18][19]. *pspc1* makes interaction with NONO via the coiled-coil domain. The *pspc1* and NONO heterodimers are essential structural components of paraspeckles, where NONO and *pspc1* proteins have more than 70% sequence identity. PSPC1 and NONO have dynamic processes that can readily exchange interaction partners to form dimerization states and change their function. This alteration is dependent on cell expression, for example, mouse Sertoli cells have high expression of SFPQ and PSPC1 compared to NONO, in contrast HeLa cells NONO and SFPQ are more abundant than PSPC1 [17]. All three DBHS members, NONO, PSPC1, and SFPQ, have 2 tandem RNA recognition motifs (RRMs) that show more than 50% sequence identity [20]. RRM motifs of *pspc1*-protein is used to target paraspeckle [1].

II. NONO PROTEIN STABILITY REGULATED BY PIN1

NONO is known as nuclear RNA- and DNA-binding protein which is a multifunctional DBHS (Drosophila behavior/human splicing) protein defined by N-terminal RNA recognition motifs (RRMs), protein-protein interaction NONA/paraspeckle domain (NOPS), and a C-terminal coiled-coil domain [21][22]. NONO protein is mainly found in the nucleus, particularly in paraspeckles [23][8], which is involved in every step of gene regulation: repression and transcriptional activation [24], termination of transcription, RNA transport [25], pre-mRNA splicing [24][26] and nuclear retention of defective RNA

for editing [27][5]. It has shown that NONO's interaction with splicing factor proline and glutamine-rich (SFPQ) regulates the DNA repair mechanism[28]. Because NONO plays a significant role in multiple processes, it is dysregulated in many cancers. For example, in breast cancer, the expression of NONO promotes the transcriptional activation of lipogenic genes and lipid production by interacting with and stabilizing the sterol regulatory element-binding protein 1 (SREBP1) [29]. In hepatocarcinoma, NONO contributes to carcinogenesis via oncogenic splicing of BIN1 protein. The BIN1 protein is involved in endocytosis and the self-destruction of cells (apoptosis)[30]. NONO is robustly expressed in prostate cancers, and promotes castration-resistant prostate cancer development by causing differential EPHA6 splicing [31][32]. Additionally, NONO expression is remarkably raised in Esophageal squamous cell carcinoma and melanoma[33], and gastric cancer cells [34], and the increased expression of NONO is associated with the aggressiveness of cancers. Peptidyl-prolyl cis-trans isomerase NIMA-interacting 1 (PIN1) regulates the conformational changes of pSer/Thr-Pro motifs and causes the change in the function, structure, or stability of various proteins[35][36]. Studies have revealed overexpression of PIN1 speeds up genomic instability and promote tumorigenesis by disrupting cell cycle coordination [37]. PIN1 is commonly dysregulated in various cancer, and its overexpression and/or overactivation is linked to a poor clinical prognosis [38], [39]. Overactivation of PIN1 agitates the balance between oncogenic and tumor-suppressing proteins, moving it towards oncogenesis; expected that more than 40 oncogenes are activated, and over 20 tumor suppressors are inactivated in numerous cancers [40]. NONO is ranked among the top proteins with similar functions to peptidylprolyl (PIN1) [41].

III. NONO INTERACTS WITH PIN1 ON ITS N-TERMINAL (WW) DOMAIN

PIN1, which is an oncoprotein, its function defined by its two domains: WW domain and PPIase domain. It has been demonstrated that the WW domain has a stronger affinity for its substrate than the PPIase domain[42] and the binding of the WW domain with the substrate enables PIN1 to perform molecular functions via the PPIase domain [43]. It is determined that PIN1 promotes the stability of NONO protein by binding at the WW domain and preventing proteasomal degradation of NONO[44].

IV. SUPT5H AND PIN1 INTERACTION IN BREAST CANCER

Aberrant expression of PIN1 is associated with the wider signaling network's distortions that fuel cancer progression. In cancers, overactivation of PIN1 is associated with poor patient survival. Because silencing of PIN1 is associated with the inhibition of breast cancer cell proliferation and tumorigenesis of breast cancer stem cells, molecular therapy for targeting PIN1 is growing [45][46]. SUPT5H is ranked among the top-ranked protein with identical functions to peptidylprolyl cis/trans isomerase (PIN1). PIN1 is a 163 residue protein containing the N-

terminal WW and C-terminal PPIase domains. Studies show that the PINNONO1 WW domain physically interacts with SUPT5H protein. PIN1 stabilizes SUPT5H and has a positive correlation[47]. SUPT5H contributes to breast cancer cell proliferation, cell cycle, invasion, migration, and apoptosis. A study revealed that the oncogenic function of SUPT5H could be a potential therapeutic target of interest to counter breast cancer [47].

V. NONO STABLES LIPID METABOLISM IN BREAST CANCER

NONO is highly expressed, approximately 63% of breast cancer[29]. Sterol regulatory-element binding proteins (SREBPs) family, including SREBP-1a, SREBP-1c, and SREBP-2 in mammalian cells, can activate the transcription of genes encoding key enzymes responsible for the biosynthesis of fatty acid and cholesterol [29]. SREBPs are master transcription factors that control lipid metabolism[48]. Studies show that endogenous NONO is associated with endogenous nSREBP-1a in HEK293T cells; this was approved by co-transfection of the Cell with NONO-GFP and nSREBP-1a-cherry, where prominent yellow color in the microscopic overlay images was detected suggesting that these two proteins interact and demonstrate physical interaction with each other. It was also shown that the conserved Y267 residue of NONO is essential for binding to the SREBP-1a protein. This conserved Y267 residue plays a considerable role in interaction with PSPC-1 too. NONO regulates the abundance of nSREBP-1a; interestingly, overexpression of NONO-WT in HEK293T cells resulted in a significant accumulation of endogenous nSREBP-1 protein.

Conversely, knockdown of the NONO gene with specific siRNA within MCF-7, HeLa cancer cells had no effects on the mRNA levels of SREBP-1a but decreased the nSREBP-1 protein. Therefore, NONO gives stability to the nSREBP-1 at the protein level [29]. NONO-WT was treated with cycloheximide, an inhibitor of protein synthesis, to examine the degradation rate of nSREBP-1, showing that degradation is significantly slower in a cell transfected with NONO-WT as compared with those transfected with vector. It is well-known that nSREBP-1a proteins are degraded through the ubiquitination-proteasome mechanism. Still, in the cancer cell, the degradation of nSREBP-1a is slow or not, which takes stability from NONO protein. Knockdown of NONO in HEK293T cells caused significantly increased ubiquitination in the over-expressed nSREBP-1a protein [29]. SREBP-1 expression is correlated with tumor size in human breast cancer-triggered tumors[48]. It was found by knockdown of the NONO gene that the lipid droplet in MCF-7 cells was decreased [29]. NONO binding to nSREBP-1a is required for SREBP-1a-dependent lipogenic gene expression. NONO promotes breast cancer cell proliferation and tumor growth through SREBP-1a. Knockdown of NONO inhibits the growth of MCF-7; however, knockdown of DBHS protein does not affect the growth of MCF-7 cells.

VI. NONO/P54 NRBAND P54-1 STRUCTURE IN DBHS COMPLEX

70% of domain sequences of NONO and P54-1 are similar; each of them has four domains for binding to RNA/DNA and protein; from the n-terminal, the domain has started in NONO from G-rich, RRM1(118-187), RRM2 (188-316), NOPS (268-316), Coiled-coil (317-375), and GP-rich .and for P54-1 from n-terminal; AP-rich,

RRM1 (84-154), RRM2 (155-234), NOPS (235-283), COILED-COIL (284-381). NONO is found in the nucleus of most mammalian cells and is primarily distributed within the nucleolus. NONO gene location is on the plus strand of the X-chromosome [49]. It can also be concentrated within sub-nuclear domains known as paraspeckles [50]. NONO has also been observed within the brain, localized in the cytoplasm of hippocampal neurons associated with RNA transport granules[49].

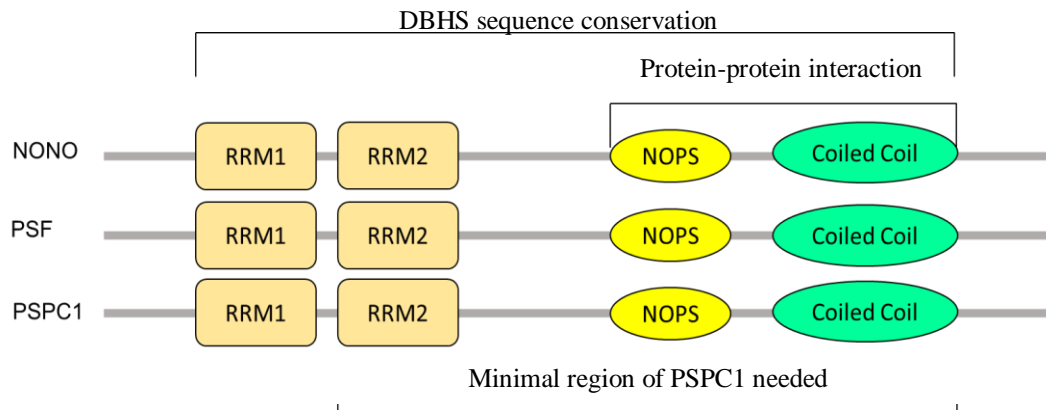


Fig. 1: The DBHS protein family shows domain structure and indicates regions involved in paraspeckle biology

VII. ROLES OF DBHS IN TRANSCRIPTIONAL REGULATION AND REPRESSION

DBHS proteins mediate transcriptional regulation and repression[16][20]. DBHS proteins associate synergistically with several transcription factors, DNA and RNA, acting bifunctionally as positive and negative transcriptional regulators. Thus, as with many transcription factors, their precise role is context-dependent[51]. The DBHS family complex act as a repressor and appears primarily driven by and dependent on SFPQ, either in a homodimer or heterodimer context. Studies have shown that SFPQ binds directly to the target gene promoter and recruits silencers such as Sin3A and HDAC[52][3]SFPQ/NONO can act on hormone receptors such as thyroid and retinoid X receptors [53] or, in complex with steroidogenic factor 1, repress the human CYP17 gene or gene involved in circadian rhythms [54][55]. SFPQ/NONO can also negatively regulate transcription by sequestering activators away from the target gene promoter. For example, direct binding of SFPQ/NONO to the progesterone receptor can prevent its binding to DNA [3].

VIII. POST-TRANSLATIONAL MODIFICATION OF DBHS

DBHS are substrates for several post-translational modifications. NONO is phosphorylated in the region proximal to the coiled-coil domain during mitosis; thus, phosphorylation of the motif region provides a binding site for PIN1 that may lead to subsequent conformational changes in this region. NONO N-terminal is also phosphorylated by CDK1 during mitosis for RNA binding to simple substrates in vitro; also, N-terminus 53 residues of NONO may allosterically regulate RNA-binding ability. In

addition, NONO is a substrate for protein phosphates, which associates with NONO RRM1 and influences NONO post-translation splicing[49].

IX. SILENCING THE NONO GENE CAN AFFECT CANCER CELL VIABILITY AND CELL CYCLE

Silencing of NONO gene expression decreases cell viability and colony-forming ability, induces apoptosis, promotes mitochondrial membrane potential changes, and reduces breast cancer cells' migratory and invasive potentials. It was demonstrated that siRNA-NONO efficiently reduced the mRNA and protein levels of the NONO gene in both MDA-MB-231 and MCF-7 cells[44]. Knockdown of the NONO gene affected on cell cycle as analyzed by flow cytometry. Knockdown of NONO in MDA-MD231 and MCF-7cell significantly arrests Cell at the S-phase of the cell cycle, at the same time, decrease the percentage of cells in the G2/M phase in the case of MDA-MB-231 and G0/G1 and G2/M phase in MCF-7 cells [44].

A. Therapeutic Targets

Mutations and dysfunction in genes are the dominant reason for the increased number of cancer cases. In the case of cancer healing, chemotherapy mechanisms are prominently used; however, targeting the specific oncologic gene by modern genome editing techniques can be a potential therapeutic approach. NONO, PIN1, and SUPT5 were significant for cancer growth, and they could be potential targets and markers for chemotherapy and gene therapy mechanisms for suppressing cancer cells [44][47]

B. Future Perspective

DBHS proteins as multifunctional nuclear proteins implicated in subnuclear body formation, co-activation, transcription initiation[2], and corepression; constitutive and alternative splicing [4]transcriptional termination[8] and DNA repair[9]. And tumor suppression[12]. DBHS complex consists of NONO, PSPC1, and SFPQ proteins, each of which has a different domain. NONO gene was found to develop breast cancer [17], prostate cancer [19], and hepatic cancer [46]. NONO gene is involved in various processes, including RNA splicing and editing, DNA unwinding and repairing, and gene transcription. Structurally, p54nrb(NONO) contains several defined domains, including RNA recognition motif domains, DNA binding domain, and a coiled-coil domain, And is involved in multifunction Beyond the interaction of NONO with paraspeckle and also interacts with PIN1 protein. Overexpression and overactivation of PIN1 accelerate genomic instability, promotes tumorigenesis by disrupting cell cycle coordination, and disrupt the balance between oncogenic and tumor-suppressing proteins, moving it towards oncogenesis; more than 40 oncogenes are activated, and over 20 tumor suppressors are inactivated in numerous cancers[40][38]. The essential role of these proteins in tumor growth is unknown. PIN1 oncoprotein is defined by its two domains: WW and PPIase domains. The NONO gene was interacted into the WW domain of PIN1 to stabilize the NONO protein. At the same time, other oncogenic proteins SUPT5H also interacted with the WW domain of PIN1 to stabilize the SUPT5H, which has a significant role in tumor progression. Here is a question how does the PIN1 protein interact with NONO and SUPT5H in the same domain? Does it any competition between NONO and SUPT5H, and how are both proteins regulated by the PIN1 protein? Indeed DBHS complex proteins are found, as a hetero-dimer or alone, in several nuclear complexes involved in the binding and processing of nucleic acids. How are these multifunctional proteins operating in the simultaneous processes occurring in the cell nucleus? Due to their dual RNA/DNA-binding properties and the ability to interact with several protein counterparts, these proteins might be regarded as 'sticky' proteins that can link multiple nuclear processes. Silencing NONO, PIN1, and SUPT5H genes suppressed the tumor's growth, indicating that these proteins will be therapeutic targets and markers for the cancerous Cell.

X. RESULT

These paraspeckle nuclear bodies are complex of three proteins, and a non-coding RNA is involved in many interaction types. DBHS consists of three proteins: NONO, PSPC1, and FSP; these proteins have a specific multidomain for binding to proteins and RNA/DNA. DBHS proteins as multifunctional nuclear proteins implicated in subnuclear body formation; co-activation, transcription initiation[2][3] and corepression; constitutive and alternative splicing[4][5][6][7] transcriptional termination[8] and DNA repair among this three protein which more has involved or to distinct interaction are NONO, pspc1 such as transcription factor, polymerase, nuclear-receptor, and gene regulation transcriptional regulation, RNA processing, transport DNA

repair mechanism, or controlling gene expression during many cellular processes including differentiation stress responses. In most cancerous cases, the expression of NONO and related proteins will increase, such as in breast cancer and malignant melanoma. NONO interacts and gives stability to nSREBP-a1, which the SREBP (sterol regulatory element-binding protein) is the protein that regulates lipogenic genes and cholesterol biosynthetic genes. NONO regulates the abundance of nSREBP-1a; interestingly, overexpression of NONO-WT in HEK293T cells resulted in a significant accumulation of endogenous nSREBP-1 protein. NONO and SUPT5H are oncogenic proteins, PIN1 and SUPT5H are both interact with WW domain of PIN1 protein, which PIN1 stabilized the NONO and SUPT5H protein. Silencing of NONO, PIN1, and SUPT5H were demonstrated the significant decrease of tumor growths.

XI. CONCLUSION

Paraspeckle are sub-nuclear bodies found in the interchromatin space of mammalian cells, made from the interaction of non-coding RNA and DBHS (Drosophila behavior human splicing) complex. DBHS, a multifunctional complex, consists of three proteins: NONO, PSPC1, and FSP; these proteins have a specific multidomain for binding to proteins and RNA/DNA. NONO gene plays a crucial role in cancer progression by involving various biological processes, including RNA splicing and editing, DNA unwinding and repairing, and gene transcription. NONO can interact with SFPQ and PSPC1 in the DBHS complex. In addition, NONO can also interact with (SREBPs) and stabilize them. (SREBPs) activate genes encoding key enzymes responsible for the biosynthesis of fatty acid and cholesterol in breast cancer. PIN1 and SUPT5H are oncogenic proteins where PIN1 interacts and stabilizes the NONO, and SUPT5H, contributing to breast cancer cell proliferation, migration, invasion, cell cycle, and apoptosis. Silencing NONO, PIN1 and SUPT5H were shown to decrease tumor growth significantly, and they can be a good target for therapeutic purposes.

ACKNOWLEDGMENTS

We thank fully the Research department of Spinghar Medical Institute Kabul Campus for their contribution in providing facilities. This article dedicated to all afghan girls deprived of education in Afghanistan.

CONFLICT OF INTEREST

We declare that we have no conflict of interest.

REFERENCES

- [1.] Y. C. Lin, L. Q. Ge, and H. J. Jiang, "Pilot study on the technique of copper and cobalt ore fast analyzing by x-ray fluorescence," *Wutan Huatan Jisuan Jishu*, vol. 29, no. 3, pp. 256–259, 2007.
- [2.] S. Kuwahara *et al.*, "PSPC1, NONO, and SFPQ are expressed in mouse sertoli cells and may function as coregulators of androgen receptor-mediated transcription," *Biol. Reprod.*, vol. 75, no. 3, pp. 352–359, 2006, doi: 10.1095/biolreprod.106.051136.
- [3.] X. Dong, O. Shylnova, J. R. G. Challis, and S. J. Lye, "Identification and characterization of the protein-associated splicing factor as a negative co-regulator of the progesterone receptor," *J. Biol. Chem.*, vol. 280, no. 14, pp. 13329–13340, Apr. 2005, doi: 10.1074/JBC.M409187200.
- [4.] J. G. Patton, E. B. Porro, J. Galceran, P. Tempst, and B. Nadal-Ginard, "Cloning and characterization of PSF, a novel pre-mRNA splicing factor," *Genes Dev.*, vol. 7, no. 3, pp. 393–406, 1993, doi: 10.1101/gad.7.3.393.
- [5.] R. Peng, B. Dye, I. Pérez, D. Barnard, A. T.- Rna, and undefined 2002, "PSF and p54nrb bind a conserved stem in U5 snRNA," *cambridge.org*, Accessed: Oct. 18, 2022. [Online]. Available: <https://www.cambridge.org/core/journals/rna/article/pf-sf-and-p54nrb-bind-a-conserved-stem-in-u5-snrna/6CDBA2EA62939282AD313D237AE865B7>.
- [6.] S. Kameoka, P. Duque, and M. M. Konarska, "P54Nrb Associates With the 5' Splice Site Within Large Transcription/Splicing Complexes," *EMBO J.*, vol. 23, no. 8, pp. 1782–1791, 2004, doi: 10.1038/sj.emboj.7600187.
- [7.] Ito *et al.*, "Brm transactivates the telomerase reverse transcriptase (TERT) gene and modulates the splicing patterns of its transcripts in concert with p54nrb," *Biochem. J.*, vol. 411, no. 1, pp. 201–209, Apr. 2008, doi: 10.1042/BJ20071075.
- [8.] S. Kaneko, O. Rozenblatt-Rosen, M. Meyerson, and J. L. Manley, "The multifunctional protein p54nrb/PSF recruits the exonuclease XRN2 to facilitate pre-mRNA 3' processing and transcription termination," *Genes Dev.*, vol. 21, no. 14, pp. 1779–1789, Jul. 2007, doi: 10.1101/GAD.1565207.
- [9.] Y. Shav-Tal and D. Zipori, "PSF and p54nrb/NonO - Multifunctional nuclear proteins," *FEBS Lett.*, vol. 531, no. 2, pp. 109–114, 2002, doi: 10.1016/S0014-5793(02)03447-6.
- [10.] S. A. Brown *et al.*, "Cell biology: PERIOD1-associated proteins modulate the negative limb of the mammalian circadian oscillator," *Science (80-.)*, vol. 308, no. 5722, pp. 693–696, 2005, doi: 10.1126/science.1107373.
- [11.] H. A. Duong, M. S. Robles, D. Knutti, and C. J. Weitz, "A molecular mechanism for circadian clock negative feedback," *Science (80-.)*, vol. 332, no. 6036, pp. 1436–1439, 2011, doi: 10.1126/science.1196766.
- [12.] G. Wang, Y. Cui, G. Zhang, A. Garen, and X. Song, "Regulation of proto-oncogene transcription, cell proliferation, and tumorigenesis in mice by PSF protein and a VL30 noncoding RNA," *Proc. Natl. Acad. Sci. U. S. A.*, vol. 106, no. 39, pp. 16794–16798, 2009, doi: 10.1073/pnas.0909022106.
- [13.] R. Stanewsky, K. G. Rendahl, M. Dill, and H. Saumweber, "Genetic and molecular analysis of the X chromosomal region 14B17-14C4 in *Drosophila melanogaster*: Loss of function in NONA, a nuclear protein common to many cell types, results in specific physiological and behavioral defects," *Genetics*, vol. 135, no. 2, pp. 419–442, 1993, doi: 10.1093/genetics/135.2.419.
- [14.] S. Nakagawa, T. Yamazaki, and T. Hirose, "Molecular dissection of nuclear paraspeckles: Towards understanding the emerging world of the RNP milieu," *Open Biol.*, vol. 8, no. 10, 2018, doi: 10.1098/rsob.180150.
- [15.] D. M. Passon, M. Lee, A. H. Fox, and C. S. Bond, "Crystallization of a paraspeckle protein PSPC1-NONO heterodimer," *Acta Crystallogr. Sect. F Struct. Biol. Cryst. Commun.*, vol. 67, no. 10, pp. 1231–1234, 2011, doi: 10.1107/S1744309111026212.
- [16.] B. Dong, D. S. Horowitz, R. Kobayashi, and A. R. Krainer, "Purification and cDNA cloning of HeLa cell p54nrb, a nuclear protein with two RNA recognition motifs and extensive homology to human splicing factor PSF and *Drosophila* NONA/BJ6," *Nucleic Acids Res.*, vol. 21, no. 17, pp. 4085–4092, 1993, doi: 10.1093/nar/21.17.4085.
- [17.] G. J. Knott, C. S. Bond, and A. H. Fox, "The DBHS proteins SFPQ, NONO and PSPC1: A multipurpose molecular scaffold," *Nucleic Acids Res.*, vol. 44, no. 9, pp. 3989–4004, 2016, doi: 10.1093/nar/gkw271.
- [18.] A. H. Fox, C. S. Bond, and A. I. Lamond, "P54nrb forms a heterodimer with PSP1 that localizes to paraspeckles in an RNA-dependent manner," *Mol. Biol. Cell*, vol. 16, no. 11, pp. 5304–5315, Nov. 2005, doi: 10.1091/MBE.E05-06-0587.
- [19.] Y. T. F. Sasaki, T. Ideue, M. Sano, T. Mituyama, and T. Hirose, "MENε/β noncoding RNAs are essential for structural integrity of nuclear paraspeckles," *Proc. Natl. Acad. Sci. U. S. A.*, vol. 106, no. 8, pp. 2525–2530, 2009, doi: 10.1073/pnas.0807899106.
- [20.] C. S. Bond and A. H. Fox, "Paraspeckles: nuclear bodies built on long noncoding RNA," *J. Cell Biol.*, vol. 186, no. 5, p. 637, Sep. 2009, doi: 10.1083/JCB.200906113.
- [21.] B. Dong, D. S. Horowitz, R. Kobayashi, and A. R. Krainer, "Purification and cDNA cloning of HeLa cell p54nrb, a nuclear protein with two RNA recognition motifs and extensive homology to human splicing factor PSF and *Drosophila* NONA/BJ6," *Nucleic Acids Res.*, vol. 21, no. 17, pp. 4085–4092, Aug. 1993, doi: 10.1093/NAR/21.17.4085.
- [22.] G. Daubner, A. Cléry, F. A.-C. opinion in structural biology, and undefined 2013, "RRM-RNA recognition: NMR or crystallography and new findings," *Elsevier*, Accessed: Oct. 18, 2022. [Online]. Available: <https://www.sciencedirect.com/science/article/pii/S0959440X1200190X>.

- [23.] C. M. Clemson *et al.*, “An Architectural Role for a Nuclear Noncoding RNA: NEAT1 RNA Is Essential for the Structure of Paraspeckles,” *Mol. Cell*, vol. 33, no. 6, pp. 717–726, Mar. 2009, doi: 10.1016/J.MOLCEL.2009.01.026.
- [24.] X. Dong, C. Yu, O. Shynlova, J. R. G. Challis, P. S. Rennie, and S. J. Lye, “p54nrb Is a Transcriptional Corepressor of the Progesterone Receptor that Modulates Transcription of the Labor-Associated Gene, Connexin 43 (Gja1),” *Mol. Endocrinol.*, vol. 23, no. 8, p. 1147, Aug. 2009, doi: 10.1210/ME.2008-0357.
- [25.] Y. Kanai, N. Dohmae, and N. Hirokawa, “Kinesin transports RNA: Isolation and characterization of an RNA-transporting granule,” *Neuron*, vol. 43, no. 4, pp. 513–525, Aug. 2004, doi: 10.1016/j.neuron.2004.07.022.
- [26.] A. Basu, B. Dong, A. R. Krainer, and C. C. Howe, “The intracisternal A-particle proximal enhancer-binding protein activates transcription and is identical to the RNA- and DNA-binding protein p54nrb/NonO,” *Mol. Cell. Biol.*, vol. 17, no. 2, pp. 677–686, Feb. 1997, doi: 10.1128/MCB.17.2.677.
- [27.] K. V. Prasanth *et al.*, “Regulating gene expression through RNA nuclear retention,” *Cell*, vol. 123, no. 2, pp. 249–263, Oct. 2005, doi: 10.1016/J.CELL.2005.08.033.
- [28.] L. Jaafar, Z. Li, S. Li, W. D.-N. acids research, and undefined 2017, “SFPQ• NONO and XLF function separately and together to promote DNA double-strand break repair via canonical nonhomologous end joining,” *academic.oup.com*, Accessed: Oct. 18, 2022. [Online]. Available: <https://academic.oup.com/nar/article-abstract/45/4/1848/2638400>.
- [29.] Z. Zhu *et al.*, “P54 nrb/NONO regulates lipid metabolism and breast cancer growth through SREBP-1A,” *Oncogene*, vol. 35, no. 11, pp. 1399–1410, 2016, doi: 10.1038/onc.2015.197.
- [30.] R. Wechsler-Reya, D. Sakamuro, J. Zhang, J. Duhadaway, and G. C. Prendergast, “Structural analysis of the human BIN1 gene: Evidence for tissue-specific transcriptional regulation and alternate RNA splicing,” *J. Biol. Chem.*, vol. 272, no. 50, pp. 31453–31458, Dec. 1997, doi: 10.1074/jbc.272.50.31453.
- [31.] H. Ishiguro, H. Uemura, K. Fujinami, N. Ikeda, S. Ohta, and Y. Kubota, “55 kDa nuclear matrix protein (nmt55) mRNA is expressed in human prostate cancer tissue and is associated with the androgen receptor,” *Int. J. cancer*, vol. 105, no. 1, pp. 26–32, May 2003, doi: 10.1002/IJC.11021.
- [32.] R. Yamamoto *et al.*, “Overexpression of p54nrb/NONO induces differential EPHA6 splicing and contributes to castration-resistant prostate cancer growth,” *Oncotarget*, vol. 9, no. 12, p. 10510, Feb. 2018, doi: 10.18632/ONCOTARGET.24063.
- [33.] S. Schiffrer, N. Zimara, R. Schmid, and A. K. Bosserhoff, “p54nrb is a new regulator of progression of malignant melanoma,” *Carcinogenesis*, vol. 32, no. 8, pp. 1176–1182, Aug. 2011, doi: 10.1093/CARCIN/BGR103.
- [34.] D. Li *et al.*, “Ets-1 promoter-associated noncoding RNA regulates the NONO/ERG/Ets-1 axis to drive gastric cancer progression,” *Oncogene 2018 3735*, vol. 37, no. 35, pp. 4871–4886, May 2018, doi: 10.1038/s41388-018-0302-4.
- [35.] E. S. Yeh and A. R. Means, “PIN1, the cell cycle and cancer,” *Nat. Rev. Cancer 2007 75*, vol. 7, no. 5, pp. 381–388, Apr. 2007, doi: 10.1038/nrc2107.
- [36.] Y. C. Liou, X. Z. Zhou, and K. P. Lu, “Prolyl isomerase Pin1 as a molecular switch to determine the fate of phosphoproteins,” *Trends Biochem. Sci.*, vol. 36, no. 10, pp. 501–514, Oct. 2011, doi: 10.1016/J.TIBS.2011.07.001.
- [37.] F. Suizu, A. Ryo, G. Wulf, J. Lim, and K. P. Lu, “Pin1 Regulates Centrosome Duplication, and Its Overexpression Induces Centrosome Amplification, Chromosome Instability, and Oncogenesis,” *Mol. Cell. Biol.*, vol. 26, no. 4, p. 1463, Feb. 2006, doi: 10.1128/MCB.26.4.1463-1479.2006.
- [38.] Z. Lu and T. Hunter, “Prolyl isomerase Pin1 in cancer,” *Cell Res. 2014 249*, vol. 24, no. 9, pp. 1033–1049, Aug. 2014, doi: 10.1038/cr.2014.109.
- [39.] T. H. Lee *et al.*, “Death-Associated Protein Kinase 1 Phosphorylates Pin1 and Inhibits Its Prolyl Isomerase Activity and Cellular Function,” *Mol. Cell*, vol. 42, no. 2, pp. 147–159, Apr. 2011, doi: 10.1016/J.MOLCEL.2011.03.005.
- [40.] X. Z. Zhou and K. P. Lu, “The isomerase PIN1 controls numerous cancer-driving pathways and is a unique drug target,” *Nat. Rev. Cancer*, vol. 16, no. 7, pp. 463–479, Jul. 2016, doi: 10.1038/NRC.2016.49.
- [41.] Y. R. Pokharel *et al.*, “Relevance Rank Platform (RRP) for Functional Filtering of High Content Protein-Protein Interaction Data,” *Mol. Cell. Proteomics*, vol. 14, no. 12, pp. 3274–3283, Oct. 2015, doi: 10.1074/MCP.M115.050773.
- [42.] P. J. Lu, X. Z. Zhou, M. Shen, and K. P. Lu, “Function of WW Domains as Phosphoserine- or Phosphothreonine-Binding Modules,” *Science (80-.)*, vol. 283, no. 5406, pp. 1325–1328, Feb. 1999, doi: 10.1126/SCIENCE.283.5406.1325.
- [43.] C. Smet, J. M. Wieruszkeski, L. Buée, I. Landrieu, and G. Lippens, “Regulation of Pin1 peptidyl-prolyl cis/trans isomerase activity by its WW binding module on a multi-phosphorylated peptide of Tau protein,” *FEBS Lett.*, vol. 579, no. 19, pp. 4159–4164, Aug. 2005, doi: 10.1016/J.FEBSLET.2005.06.048.
- [44.] B. A. Lone, I. Sharma, and F. Ahmad, “Non-POU Domain-Containing Octamer-Binding (NONO) Protein Stability Regulated by PIN1 is Crucial for Breast Cancer Tumorigenicity Via the MAPK / β - Catenin Pathway,” 2022.
- [45.] S. K. L. Karna, F. Ahmad, B. A. Lone, and Y. R. Pokharel, “Knockdown of PTOV1 and PIN1 exhibit common phenotypic anti-cancer effects in MDA-MB-231 cells,” *PLoS One*, vol. 14, no. 5, p. e0211658, May 2019, doi: 10.1371/JOURNAL.PONE.0211658.

- [46.] A. Rustighi *et al.*, “Prolyl-isomerase Pin1 controls normal and cancer stem cells of the breast,” *EMBO Mol. Med.*, vol. 6, no. 1, pp. 99–119, Jan. 2014, doi: 10.1002/EMMM.201302909.
- [47.] B. A. Lone, F. Ahmad, S. K. L. Karna, and Y. R. Pokharel, “SUPT5H post-transcriptional silencing modulates PIN1 expression, inhibits tumorigenicity, and induces apoptosis of human breast cancer cells,” *Cell. Physiol. Biochem.*, vol. 54, no. 5, pp. 928–946, 2020, doi: 10.33594/000000279.
- [48.] D. Guo, E. H. Bell, P. Mischel, and A. Chakravarti, “Targeting SREBP-1-driven lipid metabolism to treat cancer,” *Curr. Pharm. Des.*, vol. 20, no. 15, p. 2619, Nov. 2014, doi: 10.2174/13816128113199990486.
- [49.] G. J. Knott, C. S. Bond, and A. H. Fox, “The DBHS proteins SFPQ, NONO and PSPC1: a multipurpose molecular scaffold,” *Nucleic Acids Res.*, vol. 44, no. 9, pp. 3989–4004, May 2016, doi: 10.1093/NAR/GKW271.
- [50.] A. H. Fox *et al.*, “Paraspeckles: a novel nuclear domain,” *Curr. Biol.*, vol. 12, no. 1, pp. 13–25, 2002.
- [51.] G. Stampfel, T. Kazmar, O. Frank, S. Wienerroither, F. Reiter, and A. Stark, “Transcriptional regulators form diverse groups with context-dependent regulatory functions,” *Nature*, vol. 528, no. 7580, pp. 147–151, 2015, doi: 10.1038/nature15545.
- [52.] X. Dong, C. Yu, O. Shynlova, J. R. G. Challis, P. S. Rennie, and S. J. Lye, “p54nrb Is a Transcriptional Corepressor of the Progesterone Receptor that Modulates Transcription of the Labor-Associated Gene, Connexin 43 (Gja1),” *Mol. Endocrinol.*, vol. 23, no. 8, p. 1147, Aug. 2009, doi: 10.1210/ME.2008-0357.
- [53.] M. Mathur, P. W. Tucker, and H. H. Samuels, “PSF Is a Novel Corepressor That Mediates Its Effect through Sin3A and the DNA Binding Domain of Nuclear Hormone Receptors,” *Mol. Cell. Biol.*, vol. 21, no. 7, p. 2298, Apr. 2001, doi: 10.1128/MCB.21.7.2298-2311.2001.
- [54.] M. B. Sewer, V. Q. Nguyen, C. J. Huang, P. W. Tucker, N. Kagawa, and M. R. Waterman, “Transcriptional activation of human CYP17 in H295R adrenocortical cells depends on complex formation among p54(nrb)/NonO, protein-associated splicing factor, and SF-1, a complex that also participates in repression of transcription,” *Endocrinology*, vol. 143, no. 4, pp. 1280–1290, 2002, doi: 10.1210/ENDO.143.4.8748.
- [55.] H. A. Duong, M. S. Robles, D. Knutti, and C. J. Weitz, “A molecular mechanism for circadian clock negative feedback,” *Science*, vol. 332, no. 6036, pp. 1436–1439, Jun. 2011, doi: 10.1126/SCIENCE.1196766.
- [56.] <https://journals.e-palli.com/home/index.php/ajbb/authorDashboard/submitmission/1105>