ISSN No:-2456-2165

A Review of the Ethnopharmacology, Phytochemistry and Pharmacology of *Rauwolfia tetraphylla*

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Abstract:- Human civilization was encountered by many insults in the form of diseases since the day of evolution. In ancient times, natural products were used as the main source of remedies to overcome the aforesaid situation. The availability of growing body of literature on the medicinal properties of the plants belonging to the family Apocyanaceae is grabbing attention towards its further research. Rauwolfia tetraphylla is one of the plant of this family having many therapeutic benefits holds an important position in Indian traditional system of medicine. The plant commonly known as "Devil-pepper" widely used among south Indian tribes due to their ethnobotanical significance. Hence, present article reviews about updated information on various ethnopharmacological activities of R. tetraphylla owing to their medicinal importance.

Keywords:- Rauwolfia Tetraphylla, Ethnopharmacological, Antioxidant Activity, Anticancer Activity, Reserpine

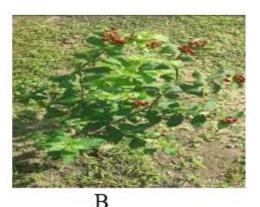
I. INTRODUCTION

Products from natural origin are estimable sources of novel structures for the ambition of drug development. Identification of plant products for their therapeutic application continues to be a growing interest from ancient time to till now [1]. Use of herbal medication is known as alternative therapy against several diseases having plentiful advantages over synthetic drugs like decreased adverse effects, cost effective, easily accessible and improved compatibility [2].

Rauwolfia teraphylla (Synonym- Rauwolfia canescens, Rauwolfia heterophylla, Rauwolfia hirsuta) commonly known as Devil pepper, Be still tree, milkbush, wild snake root, American serpentwood in english, Barachandrika, chandrabhaga in hindi, pampukaalaachchedi in tamil, pampumkolli, Kattamalpori in malayalam, Papataku in telugu, bar chandrika, gandhankuli in bengali, patalagarudi in odia and sarpanasini in sanskrit belongs to family Apocyanacae is a species facing extinction being native to West Indies and now cultivated commercially in many countries such as India, Pakistan, Sri Lanka, Bangladesh, Nepal, and Myanmar and often cultivated as garden plants. Distribution of the plant in India includes several states such as West Bengal, Karnataka, Odisha, Bihar, Andhra Pradesh, Tamilnadu and Kerala. R. tetraphylla having illustrious therapeutic potential described in ancient literature triggers the widely practice in traditional system of medicine. The plant contains alkaloids such as reserpine, rauvolscine, ajmalicine, ajmaline, canescine, pseudoyohimbine, and yohimbine. R. tetraphylla contains a number of chemicals and several studies carried out on the plant have revealed various pharmacological activities [3-15].



Fig 1:- A- Fruiting stem, B- Whole plant of Rauwolfia tetraphylla



II. BOTANICAL CHARACTERIZATION

R. tetraphylla (Figure 1) synonym *R. canescens* is a pubescent, ever-green shrub with woody stem and reaching a height of 4-6 feet. Leaves are unequal, 5-9 x 3-4 cm, elliptic-ovate, acute at apex, pubescent and usually found in whorls of 4. Flowers are cream colored, about 5mm across, found in terminal corymbose cymes. Calyx lobes are short, ciliate and round. Corolla is white in color, approximately 3mm long, lobes and tubes are short. Drupes are ovoid, 2-seeded, 5-10mm across, smooth, jointed to the top, purple when ripe. Flowering occurs throughout the year [10,16,17].

III. TRADITIONAL USES

Plants are an integral component of traditional medicine. Throughout world, whole plants or certain parts of the plants have been widely used to cure several diseases and disorders either in single or polyherbal formulations. Plant based medicines are cheaper, usually devoid of side effects (that are associated with the use of modern medicines like antibiotics and anticancer agents), confer multifold beneficial effects and are locally available [18-26].

The plant *R. tetraphylla* possesses a range of ethnobotanical applications in various countries across the world as indicated by several studies. The latex from the plant is reported to be cathartic, emetic, and expectorant besides its use in the treatment of dropsy. The juice prepared from the fruit is used as a substitute for ink. Leaf juice is used in eye troubles and decoction prepared from leaves is used for toothache. Root is reported to have antihypertensive, sedative, hypnotic effect. Paste prepared from roots is useful in stomach pain and snake bite [5,27,28,29].

R. tetraphylla is one among the various ethnomedicinal plants being used by amerindian communities of Caribbean basin [30]. The roots of the plant are used in nervous disorders and insomnia in Gopalganj district of Bihar, India [31]. Whole plant as well as different parts such as root, fruit and leaves of R. tetraphylla are widely used in traditional medicine (in various forms such as paste, powder, decoction and juice), particularly snake bite. In Bagota, Colombia, the plant is used as an antidote and for blood pressure [32].

The leaves of *R. tetraphylla* are used ethnomedicinally by Peasant community of San Jacinto, Northern Colombia to relieve tension [33]. Roots are used in high blood pressure and mental disorders in Coimbatore district, Tamilnadu, India [34]. The leaves and roots are used against nervous disorders, anxiety, excitement, cough and fever in Barpeta district, Assam, India [35]. Roots are used for treating nervous disorders in Villupuram district, Tamil Nadu, India [36].

The Nyishi tribe of Arunachal Pradesh, India uses roots of the plants as anthelmintic, antidote to snake bite and to cure vomiting, insomnia, insanity, blood pressure, malaria [37]. The leaves are used traditionally for treating eczema in Tamil Nadu, India [38]. The leaves and fruits are used against snake bite in Thiruvannamalai district, Tamil Nadu, India [39]. In Odisha. India, the leaves and roots are used to treat stomachache [40]. Roots and whole plant of R. tetraphylla are used to increase uterine contraction, as anthelmintic and to treat skin diseases and hypertension in South West Bengal, India [41]. In Midnapore district, West Bengal, India, the roots are used as sedative, hypnotic and for hypertension. Leaf juice is used for the purpose of removal of opacities of cornea of eyes [42]. The whole plant and root or R. tetraphylla are used in the treatment of impotency and to increase libido in Greater Naogaon district, Bangladesh [43]. The plant is used as an antidote to poison in Rajshahi district, Bangladesh [44]. In Chuadanga and Jhenaidah districts of Bangladesh, different parts i.e. leaf, stem and root are used in snake bite and as snake repellant [45]. In Vizianagaram district of Andhra Pradesh, India the plant is used to treat blood pressure [46]. In Chittagong, Bangladesh, the leaves and roots are used against high blood pressure [47]. Various parts such as seed, leaf and root of R. tetraphylla are used to treat snake bite in Kancheepuram district, Tamil Nadu, India [48]. The roots are used to treat mental disorders and high blood pressure in Erode district, Tamil Nadu, India [49].

IV. PHYTOCHEMISTRY

Plants produce a range of chemicals that can be divided into two categories *viz.* primary and secondary metabolites. These chemicals are known as phytochemicals and most of the secondary metabolites (with complex chemical composition), for e.g., alkaloids, terpenes and polyphenolic compounds, exert multifold effects on the health of human beings besides conferring resistance to plants that produce

ISSN No:-2456-2165

them against insects, pathogens and herbivores. Secondary metabolites in plants are restricted in distribution within the plant kingdom i.e. some metabolites are present in only one plant species or in a related group of species. Metabolic pathways such as shikimic acid pathway, malonic acid pathway and mevalonic acid pathway are responsible for synthesis of secondary metabolites in plants [50-58]. Due to advancements in chromatographic and spectral methods, a vast number of plant secondary metabolites have been identified and their structures are proposed [59-65].

R. tetraphylla is shown to contain a myriad of secondary metabolites, particularly alkaloids. Reserpine is an important alkaloid found in R. tetraphylla. With the help of chromatographic and spectral analyses, many secondary compounds have been isolated from different parts of R. tetraphylla and their structures have been elucidated. Klohs et al. [66] isolated reserpine from roots of R. tetraphylla. Stoll and Hofmann [67] isolated two alkaloids canescine and pseudovohimbine from the roots of *R. tetraphylla*. Two ester alkaloids, named as raunescine and isoraunescine, have been isolated from R. tetraphylla [68]. An alkaloid designated as Recanescine (that lack the methoxyl group in the C-ll position of reserpine) with sedative and hypotensive property was isolated from R. tetraphylla [69,70]. An alkaloid designated as heterophyllin was isolated from roots of R. tetraphylla [71]. Raunescine (renamed as canembine), an alkaloid, was isolated from roots of R. tetraphylla [72]. An alkaloid pseudoreserpine possessing hypotensive and sedative activity

was isolated from the root of *R. tetraphylla* [73]. Djerassi et al. [74] elucidated the structures of two alkaloids from root of *R. tetraphylla* i.e. tetraphylline and tetraphyllicine. Belikov [75] isolated alkaloids namely ajmaline, ajmalieine, arieine, reserpiline, α-yohimbine, sarpagine and reserpine from roots and epigeal parts of *R. tetraphylla*.

A new sarpagine-type alkaloid, named as N (α)-Demethylaccedine, was isolated from stem bark of R. tetraphylla [76]. By high-performance thin-layer chromatography (HPTLC) analysis, reserpine and ajmalicine were identified in root of R. tetraphylla [77]. Quercetin was identified in the leaves of R. tetraphylla by HPTLC analysis Alkaloids 10-methoxytetrahydroalstonine, isoreserpiline, α-yohimbine, reserpiline was isolated from chloroform fraction of R. tetraphylla leaves by pH-zonerefining fast centrifugal partition chromatography [79]. Six indole alkaloids isoreserpiline, viz. methoxytetrahydroalstonine, 11-demethoxyreserpiline, 10demethoxyreserpiline, α-vohimbine and reserpiline have been identified from leaves of R. tetraphylla [80]. A new labdane diterpene characterized as 3β-hydroxy-labda-8(17),13(14)-dien-12(15)-olide (Figure 2) was isolated from air-dried stems and branches of R. tetraphylla [81]. The study by Panda et al. [82] revealed a varying concentration of reserpine in different parts of R. tetraphylla. Root was shown to contain high concentration of reserpine followed by stem and leaf.

PHYTOCONSTITUENT	STRUCTURE	MEDICINAL USES	REFERENCE
RESERPINE	H ₃ CO ₂ C OCH ₃ H ₃ CO ₂ C OCH ₃	Sedative and hypotensive activity	Klohs et al., 1957
CANESCINE	NH H	Antihypertensive agent	Stoll and Hofmann et al., 1955
PSEUDOYOHIMBINE	н ₅ СО ОН	Antihypertensive, Treatment of prostate and aphrodisiac activity	Stoll and Hofmann et al., 1955
RAUNESCINE	H H N OCH3	Tranquilizing activity	Hosansky and Smith et al., 1955
RECANESCINE	H ₃ C CH ₃	Antihypertensive and sedative	Slater et al., 1955 Neuss et al., 1955

HETEROPHYLLIN	HE TO SEE THE	Antiplasmodial, antitrypanosomal, and cytotoxic activities	Hochstein et al., 1955
TETRAPHYLLINE	R O OH OH	Antianxiety activity	Djerassi et al., 1957
TETRAPHYLLICINE	HO, HH	Central tranquilizing and prolonged hypotensive activity	Djerassi et al., 1957
AJMALINE	HO N HO OH	Antiarrhythmic activity	Belikov et al., 1969
AJMALICINE	N H H	Antihypertensive activity	Belikov et al., 1969
RESERPILINE	H ₃ CO H H CH ₃	Hypotensive and antimicrobial activity	Belikov et al., 1969
A-YOHIMBINE	H ₃ CO OH	Increase libido and antianxiety activity	Belikov et al., 1969
SARPAGINE	но н он	Antiarrythmic, antihypertensive and antimicrobial activity	Martinez et al., 1989

Table 1:- Phytoconstituents and their uses of R. Tetraphylla

V. PHARMACOLOGICAL ACTIVITIES

Due to the presence of diversified phytoconstituents (most importantly alkaloids) *R. tetraphylla* acquires several distinctive pharmacological properties. Numerous reported activities are listed below.

➤ Antimicrobial Activity:

A study was conducted on petroleum ether, ethyl acetate and chloroform extracts of dry fruits of the plant for determining antibacterial activity using agar diffusion method against eight bacterial species. The results indicated that the dry fruit extracts exhibited potential broad spectrum antibacterial activity [83].

The *In-vitro* antibacterial activity of ethanolic extracts of *R. tetraphylla* was assessed by disc diffusion method against pathogenic bacteria. The results indicated that inhibitory activity was comparable with standard antibiotic (ciprofloxacin) [84]. *R. tetraphylla* calli extract and leaves were tested for *In-vitro* antimicrobial activity using broth dilution method against pathogenic fungi and bacteria. The calli extract and leaves were extracted by petroleum ether, chloroform, benzene, methanol and absolute alcohol. The results indicated that the methanolic extract exhibited MIC of 0.5 to 100 mg/ml against pathogenic fungi and against bacterial pathogens have shown MIC of about 0.25 to 100 mg/ml [85]. The antimicrobial activity of *R. tetraphylla* ethanolic extract was determined against various fungi and bacteria. It was observed that the extract exhibited good

ISSN No:-2456-2165

antimicrobial activity against A.niger, E.aerogenes and E.coli [86].

➤ Anti-inflammatory Activity

Anti-inflammatory activity was studied by orally administering various bark extracts of *R. tetraphylla* root using Carrageenan induced acute inflammation in rats and results have indicated good activity [87].

➤ Anticancer Activity

A study reported isolation and structure elucidation of 3β -hydroxy-labda-8(17),13(14)-dien-12(15)-olide, new labdane diterpene. It was tested for anticancer and antitumor activity against breast cancer cell lines such as Michigan Cancer Foundation-7 (MCF-7), basal breast cancer cell line MDA-MB-468 and claudin-low cell line MDA-MB-231, human cancer cell lines such as KB cells a sub line of the ubiquitous KERATIN-forming tumor cell line HeLa, Human epithelial type 2 (Hep-2) cells, A549 and leukemic T cells. The labdane diterpene were found to show significant activity [88].

➤ Anti-psychotic Activity

The methanolic extracts of *R.tetraphylla* leaves were prepared and assessed for antipsychotic activity using female Swiss albino mice [89]. The antipsychotic activity was determined by amphetamine induced hyperactive mouse model in-vivo and against serotogenic (5-HT2A) and dopaminergic (DA-D2) receptors *in-vitro* [90]. Further safety and toxicity evaluation studies of *R. tetraphylla* leaves methanolic extracts on female Swiss albino mice at different doses (10, 100, 300 and 2000 mg/kg) and results indicated that methanolic extract is non toxic. The isolated alkaloids might serve as a promising lead structure in drug development for treating psychotic conditions in human [91].

➤ Antioxidant Activity

The antioxidant potential of five different species of Rauwolfia leaves (R. serpentine, R. densiflora, R. tetraphylla, R. beddomei and R. micrantha) from Western Ghats was investigated. The antioxidant activity was assessed using various in-vitro models like reducing power, 1, 1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity, total antioxidant capacity, and superoxide anion scavenging activity at different concentrations. R. tetraphylla have shown the highest concentration of Lycopene and β -carotene [92].

➤ Antihypertensive Activity

The roots were found to be useful in the treatment of cardiovascular diseases and hypertension [93].

➤ Miscellaneous Activities

The extract of *R. tetraphylla leaves* is useful in the treatment of diarrhoea, dysentery, cholera, eye disease, intestinal disorders and fever [94]. The decoction of the plant bark is useful in treatment of skin disease, chronic refractory and for destroying parasites whereas, the plant extract when mixed with castor oil is used for treatment of refractory ailments [95]. The roots are recommended to be used in resolving difficult cases of child birth as they induce uterine contraction [96].

VI. CONCLUSION

The traditional herbal medicine is considered as a prominent health care system and used for primary treatment in the developing countries. R. tetraphylla chemically reported to possess diverse group of bioactive compounds like alkaloids, flavonoids, phenols, tannins, terpenes and triterpenes. The plant has been used for therapy of the various ailments and the presence of above bioactive compounds in the plant can be attributed to their medicinal values. The various plant part extracts have been scientifically proven to the pharmacological activities namely Antimicrobial, anti-inflammatory, antipsychotic, anticancer, antihypertensive, treatment of cholera, intestinal disorders, diarrhoea, dysentery, eye disease, fever and skin diseases. Hence, further research should be initiated in the isolation, characterization and pharmacological screening of isolated pure bioactive components of R. tetraphylla.

ACKNOWLEDGEMENT

The authors are contented to the department of Pharmacology of Faculty of Pharmaceutical Sciences, Siksha 'O' Anusandhan (SOA) Deemed to be University for rendering adequate facility in the laboratory for the purpose of this research and also to Dr. Manoj Ranjan Nayak for his kind support and cooperation.

REFERENCES

- [1]. Newman, D.J., Cragg, G.M., 2012. Natural products as sources of new drugs over the 30 years from 1981 to 2010. J. Nat. Prod. 75, 311–335. https://doi.org/10.1021/np200906s.
- [2]. Yadav, N.K., Arya, R.K., Dev, K., Sharma, C., Hossain, Z., Meena, S., Arya, K.R., Gayen, J.R., Datta, D., Singh, R.K., 2017. Alcoholic extract of Eclipta alba shows *invitro* antioxidant and anticancer activity without exhibiting toxicological effects. Oxid.Med. Cell Longev. 1–18. 2017. https://doi.org/10.1155/2017/9094641.
- [3]. "Rauvolfia tetraphylla Wild Snake Root", *Flowersofindia.net*, 2019. [Online]. Available: http://www.flowersofindia.net/catalog/slides/Wild%20 Snake%20Root.html. [Accessed: 06- Nov- 2019]
- [4]. Srivastava A, Tripathi AK, Pandey R, Verma RK, Gupta MM. Quantitative determination of reserpine, ajmaline, and ajmalicine in Rauvolfia serpentina by reversed phase high-performance liquid chromatography. Journal of Chromatographic Science 2006; 44: 557-560.
- [5]. Khare CP. Indian Medicinal Plants. An Illustrated Dictionary. Springer-Verlag New York, 2007, Pp 539.
- [6]. Faisal M, Alatar AA, Ahmad N, Anis M, Hegazy AK. An efficient and reproducible method for in-vitro clonal multiplication of Rauvolfia tetraphylla L. and evaluation of genetic stability using DNA-based markers. Appl Biochem Biotechnol 2012; 168: 1739-1752.

- [7]. Jyothi T, Brijesh K, Venkatesh HKR. Pharmacognostical evaluation of Rauvolfia tetraphylla L. J Pharm Sci Innovation 2012; 1(6): 57-60.
- [8]. Geng C, Liu X. Five new indole alkaloids from the leaves of Rauvolfia yunnanensis. Fitoterapia 2013; 89: 42-47.
- [9]. Anitha S, Kumari RBD. *In-vitro* callus culture in Rauvolfia tetraphylla L. Indole alkaloid production. Asian J Plant Sci 2013; 12(1): 28-33.
- [10]. Bhat GK. Flora of South Kanara, Akriti Prints, Mangalore, India, 2014, Pp 691.
- [11]. Rahman MAHM, Akter M. Taxonomy and traditional medicinal uses of Apocynaceae (Dogbane) family of Rajshahi district, Bangladesh. Research and Reviews: Journal of Botanical Sciences 2015; 4(3): 1-12.
- [12]. Gao Y, Yu A, Li G, Hai P, Li Y, Liu J, Wang F. Hexacyclic monoterpenoid indole alkaloids from Rauvolfia verticillata. Fitoterapia 2015a; 107; 44-48.
- [13]. Tamboli SR, Pandit RS. Subacute toxicity evaluation of Rauvolfia tetraphylla methanolic leaf extract in Sprague dawley rat. Int J Pharm Sci Rev Res 2015; 34(1): 152-157.
- [14]. Lobay D. Rauwolfia in the treatment of hypertension. Integrative Medicine 2015; 14(3): 40-46.
- [15]. Vaniswari DS, Shyamala K, Sivaranjani K. Adulterants and substitutes for herbal plants in Siddha system of medicine. World J Pharm Pharm Sci 2018; 7(1): 244-268.
- [16]. Kamble RB, Hate S, Chaturvedi A. New additions to the flora of Nagpur district, Maharashtra. Journal on New Biological Reports 2013; 2(1): 9-13.
- [17]. Patil MB. Ethnomedicines of Nandurbar district, Maharashtra. Laxmi book publication, Solapur, Maharashtra, India, 2015; Pp 149.
- [18]. Hutchings A. A survey and analysis of traditional medicinal plants as used by the Zulu, Xhosa and Sotho. Bothalia 1989; 19(1): 111-123.
- [19]. Sheng-Ji P. Ethnobotanical approaches of traditional medicine studies: some experiences from Asia. Pharm Biol 2001; 39: 74-79.
- [20]. Bussmann RW, Sharon D. Traditional medicinal plant use in Northern Peru: tracking two thousand years of healing culture. J Ethnobiol Ethnomed 2006; 2: 47.
- [21]. Liu Y, Dao Z, Yang C, Liu Y, Long C. Medicinal plants used by Tibetans in Shangri-la, Yunnan, China. J Ethnobiol Ethnomed 2009; 5: 15.
- [22]. Khongsai M, Saikia SP, Kayang H. Ethnomedicinal plants used by different tribes of Arunachal Pradesh. Indian J Tradit Know 2011; 10(3): 541-546.
- [23]. Vásquez J, Jiménez SL, Gómez IC, Rey JP, Henao AM, Marín DM, Romero JO, Alarcón JC. Snakebites and ethnobotany in the Eastern region of Antioquia, Colombia--the traditional use of plants. J Ethnopharmacol 2013; 146(2): 449-455.
- [24]. Eldeen IMS, Effendy MAW, Tengku-Muhammad TS. Ethnobotany: Challenges and future perspectives. Research Journal of Medicinal Plants 2016; 10: 382-387
- [25]. Kekuda PTR, Raghavendra HL, Bharadwaj NA, Akhilesha A. Traditional uses, chemistry and pharmacological activities of *Leea indica* (Burm. f.)

- Merr. (Vitaceae): A comprehensive review. Int J Green Pharm 2018; 12(Suppl 1): S71-S80.
- [26]. Jamshidi-Kia F, Lorigooini Z, Amini-Khoei H. Medicinal plants: Past history and future perspective. J Herbmed Pharmacol 2018; 7(1): 1-7.
- [27]. Caamal-Fuentes E, Torres-Tapia LW, Simá-Polanco P, Peraza-Sánchez SR, Moo-Puc R. Screening of plants used in Mayan traditional medicine to treat cancer-like symptoms. J Ethnopharmacol 2011; 135: 719-724.
- [28]. Quattrocchi U. CRC world dictionary of medicinal and poisonous plants. CRC press, 2012, Pp 3181.
- [29]. Choudhury S, Rahaman CH, Mandal S, Ghosh A. Folklore knowledge on medicinal usage of the tribal belts of Birbhum district, West Bengal, India. International Journal of Botany and Research 2013; 3(2): 43-50.
- [30]. Torres-Avilez W, Méndez-González M, Durán-García R, Boulogne I, Germosén-Robineau L. Medicinal plant knowledge in Caribbean Basin: a comparative study of Afrocaribbean, Amerindian and Mestizo communities. J Ethnobiol Ethnomed 2015; 11: 18.
- [31]. Prasad SK, Singh BN. Documentation of ethno medicinal plants of Gopalganj district of Bihar (India). IOSR Journal of Pharmacy and Biological Sciences 2014; 9(3): 80-89.
- [32]. Bussman RW, Zambrana NYP, Romero C, Hart RE. Astonishing diversity-the medicinal plant markets of Bogotá, Colombia. J Ethnobiol Ethnomed 2018; 14: 43.
- [33]. Bonzani RM. Medicinal use of plants by the Peasant community of San Jacinto, Northern Colombia. Caldasia 1999; 21(2): 203-218.
- [34]. Kumar JS, Kumar AR, Uma G, Subbiyan B, Aravindhan V, Balasubramaniam V. Survey and documentation of commercially sold medicinal plants in local markets of Velliangiri hills (Poondi), Coimbatore district, Tamil Nadu, India. International Journal of Recent Advances in Multidisciplinary Research 2015; 2(12): 1047-1055.
- [35]. Das R. Biodiversity of ethnomedicinal plants used by the ethnic tribal people of Barpeta district of Assam, north east India. Asian Journal of Pharmaceutical Science and Technology 2016; 6(1): 27-32.
- [36]. Arulappan TM, Britto JS, Ruckmani K, Kumar MR. An ethnobotanical study of medicinal plants used by ethnic people in Gingee hills, Villupuram district, Tamilnadu, India. American Journal of Ethnomedicine 2015; 2(2): 84-102.
- [37]. Tripathi AK, Limasenla, Shankar R. Ethno-medicinal plants used by Nyishi tribe of Arunachal Pradesh, India. World J Pharm Pharm Sci 2017; 6(5): 1246-1253.
- [38]. Latha R, Agastian P. An investigation on pharmaceutical ethnobotanicals used by the primitive tribes of five areas in the Eastern Ghats of India. World J Pharm Res 2015; 4(9): 1437-1464.
- [39]. Ranganathan R, Vijayalakshmi R, Parameswari P. Ethnomedicinal plants and their utilization by villagers in Jawadhu hills of Thiruvannamalai district of Tamilnadu, India. International Journal of Pharmaceutical Research and Development 2012; 4(4): 174-183.

- [40]. Devi U, Dwivedi H, Aminuddin, Zakir M, Khan H. Traditional phytotherapy of Jajpur forests of Eastern Ghat, Odisha. Hippocratic Journal of Unani Medicine 2016; 11(1): 101-120.
- [41]. Tripathi S, Ray S, Mondal AK, Verma NK. Rare ethno medicinal plants of south west Bengal, India with their different medicinal uses: needs conservation. Int J Life Sci Biotechnol Pharma Res 2013; 2(2): 114-122.
- [42]. Das PK, Mondal AK. A contribution to the medicinal plants of West Midnapore district, West Bengal, India. Ethnobotany and medicinal plants of India and Nepal. Volume 3. Singh V. (Editor), Scientific Publisher (India), Jodhpur, India, 2009, Pp 128-138.
- [43]. Rahmatullah M, Hasan MR, Hossan MS, Jahan R, Chowdhury MH, Seraj S, Miajee ZUMEU, Azad AK, Bashar AABM, Islam F. A Survey of medicinal plants used by folk medicinal practitioners of six villages in greater Naogaon district, Bangladesh. American-Eurasian Journal of Sustainable Agriculture 2010; 4(3): 309-325.
- [44]. Mollik MAH, Hossan MS, Paul AK, Taufiq-Ur-Rahman M, Jahan R, Rahmatullah M. Comparative analysis of medicinal plants used by folk medicinal healers in three districts of Bangladesh and inquiry as to mode of selection of medicinal plants. Ethnobotany Research & Applications 2010; 8: 195-218.
- [45]. Khatun A, Khan MAA, Rahman MA, Akter MS, Hasan A, Parvin W, Ripa RJ, Moniruzzaman M, Mahal MJ, Rahmatullah M. Ethnomedicinal usage of plants and animals by folk medicinal practitioners of three villages in Chuadanga and Jhenaidah districts, Bangladesh. American-Eurasian Journal of Sustainable Agriculture 2013; 7(5): 319-339.
- [46]. Parijatham TR, Sujatha B, Lakshmi SB. Ethnomedicinal studies of medicinal plants in Eastern ghats of Vizianagaram district, Andhra Pradesh, India. International Journal of Bioassays 2016; 5: 4825-4842.
- [47]. Biswas A, Bari MA, Roy M, Bhadra SK. Inherited folk pharmaceutical knowledge of tribal people in the Chittagong Hill tracts, Bangladesh. Indian J Tradit Know 2010; 9(1): 77-89.
- [48]. Senthilkumar K, Mathialagan P, Manivannan C. Herbal snake bite remedies of Irula tribal people of Kancheepuram district, Tamil Nadu, India. Int J Curr Microbiol Appl Sci 2018; 7(7): 425-438.
- [49]. Vijayashalini P, Abirami P. Diversity of medicinal plants in Eratti hill, Thamaraikarai beat of Bargur reserve forest, Western Ghats in Erode district, Tamilnadu, India. Asian J Pharm Clin Res 2018; 11(10): 78-85.
- [50]. Pichersky E, Gang DR. Genetics and biochemistry of secondary metabolites in plants: an evolutionary perspective. Trends in Plant Science Perspective 2000; 5(10): 439-445.
- [51]. Acamovic T, Brooker JD. Biochemistry of plant secondary metabolites and their effects in animals. Proc Nutr Soc 2005; 64: 403-412.
- [52]. Karuppusamy S. A review on trends in production of secondary metabolites from higher plants by *in-vitro* tissue, organ and cell cultures. Journal of Medicinal Plants Research 2009; 3(13): 1222-1239.

- [53]. Kennedy DO, Wightman EL. Herbal extracts and phytochemicals: Plant secondary metabolites and the enhancement of human brain function. Adv Nutr 2011; 2(1): 32-50.
- [54]. Mazid M, Khan TA, Mohammad F. Role of secondary metabolites in defense mechanisms of plants. Biology and Medicine 2011; 3(2): 232-249.
- [55]. Ribera AE, Zuñiga G. Induced plant secondary metabolites for phytopatogenic fungi control: a review. Journal of Soil Science and Plant Nutrition 2012; 12(4): 893-911.
- [56]. Kabera JN, Semana E, Mussa AR, He X. Plant secondary metabolites: biosynthesis, classification, function and pharmacological properties. J Pharm Pharmacol 2014; 2: 377-392.
- [57]. Ahmed E, Arshad M, Khan MZ, Amjad MS, Sadaf HM, Riaz I, Sabir S, Ahmad N, Sabaoon. Secondary metabolites and their multidimensional prospective in plant life. J Pharmacogn Phytochem 2017; 6(2): 205-214.
- [58]. Guerriero G, Berni R, Munoz-Sanchez AJ, Apone F, Abdel-Salam EM, Qahtan AA, Alatar AA, Cantini C, Cai G, Hausman J, Siddiqui KS, Hernandez-Sotomayor TSM, Faisal M. Production of plant secondary metabolites: Examples, tips and suggestions for biotechnologists. Genes 2018; 9: 309.
- [59]. Macias FA, Galindo JLG, Galindo JCG. Evolution and current status of ecological phytochemistry. Phytochemistry 2007; 68: 2917-2936.
- [60]. Boligon AA, Athayde ML. Importance of HPLC in analysis of plants extracts. Austin Chromatogr 2014; 1(3): 2.
- [61]. Bajpai VK, Majumder R, Park JG. Isolation and purification of plant secondary metabolites using column-chromatographic technique. Bangladesh Journal of Pharmacology 2016; 11(4): 844-848.
- [62]. Seal T. Quantitative HPLC analysis of phenolic acids, flavonoids and ascorbic acid in four different solvent extracts of two wild edible leaves, Sonchus arvensis and Oenanthe linearis of North-Eastern region in India. Journal of Applied Pharmaceutical Science 2016; 6(2): 157-166.
- [63]. Altemimi A, Lakhssassi N, Baharlouei A, Watson DG, Lightfoot DA. Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts. Plants 2017; 6: 42.
- [64]. Keskes H, Belhadj S, Jlail L, El Feki A, Damak M, Sayadi S, Allouche N. LC-MS–MS and GC-MS analyses of biologically active extracts and fractions from Tunisian *Juniperus phoenice* leaves. Pharm Biol 2017; 55(1): 88-95.
- [65]. Ingle KP, Deshmukh AG, Padole DA, Dudhare MS, Moharil MP, Khelurkar VC. Phytochemicals: Extraction methods, identification and detection of bioactive compounds from plant extracts. J Pharmacogn Phytochem 2017; 6(1): 32-36.
- [66]. Klohs MW, Draper MD, Keller F, Petracek FJ. The isolation of reserpine from *Rauwolfia canescens* Linn. J Am Chem Soc 1954; 76(5): 1381-1381.

- [67]. Stoll A, Hofmann A. Canescine and pseudoyohimbine from the roots of *Rauwolfia canescens*. J Am Chem Soc 1955; 77(3): 820–821.
- [68]. Hosansky N, Smith E. Raunescine and isoraunescine from *Rauwolfia canescens* L. Journal of the American Pharmaceutical Association 1955; 44(10): 639-640.
- [69]. Slater IH, Rathbun RC, Henderson FG, Neuss N. Pharmacological properties of recanescine, a new sedative alkaloid from *Rauwolfia canescens* Linn. Proc Soc Exp Biol Med 1955; 88(2): 293-295.
- [70]. Neuss N, Boaz HE, Forbes JW. Rauwolfia alkaloids. III.1 Recanescine, a new sedative principle of *Rauwolfia canescens* Linn. J Am Chem Soc 1955; 77(15): 4087–4090.
- [71]. Hochstein FA, Murai K, Boegemann WH. Alkaloids of *Rauwolfia heterophylla*1. J Am Chem Soc 1955; 77(13): 3551-3554.
- [72]. Bhattacharji S, Dhar MM, Dhar ML. Raunescine -A subsidiary alkaloid from *Rauvolfia canescens* Linn. Journal of Scientific and Industrial Research 1956; 15B: 506-507.
- [73]. Klohs MW, Keller F, Williams RE, Kusserow GW. Alkaloids of *Rauwolfia canescens* Linn. IV.1 The structure of pseudoreserpine. J Am Chem Soc 1957; 79(14): 3763-3766.
- [74]. Djerassi C, Fishman J, Gorman M, Kutney JP, Pakrashi SC. Alkaloid Studies. XVI.1 Alkaloids of *Rauwolfia tetraphylla* L. The Structures of Tetraphylline and Tetraphyllicine2, 3. J Am Chem Soc 1957; 79(5): 1217-1222.
- [75]. Belikov AS. Alkaloids of *Rauwolfia canescens*. Khimiya Prirodnykh Soedinenii 1969; 5(1): 64.
- [76]. Martinez JA, Velez H, Santana T. An alkaloid from two *Rauwolfia* spp. Phytochemistry 1989; 28(3): 961-962.
- [77]. Pandey DK, Radha, Dey A. A validated and densitometric HPTLC method for the simultaneous quantification of reserpine and ajmalicine in *Rauvolfia serpentina* and *Rauvolfia tetraphylla*. Brazilian Journal of Pharmacognosy 2016; 26(5): 553-557.
- [78]. Satyanarayana B, Kasturi P, Devi SP. Phytochemical and HPTLC studies on three medicinally important plants *Rauvolfia tetraphylla*, *Phyllanthus niruri* and *Carissa carandas*. International Journal of Research in Pharmacy and Chemistry 2018; 8(1): 44-51.
- [79]. Maurya A, Gupta S, Srivastava SK. Large-scale separation of antipsychotic alkaloids from *Rauwolfia tetraphylla* L. by pH-zone-refining fast centrifugal partition chromatography. Journal of Separation Science 2013; 36(2): 407-413.
- [80]. Gupta S, Khanna VK, Maurya A, Bawankule DU, Shukla RK, Pal A, Srivastava SK. Bioactivity guided isolation of antipsychotic constituents from the leaves of *Rauwolfia tetraphylla* L. Fitoterapia 2012b; 83(6): 1092-1099.
- [81]. Brahmachari G, Mandal LC, Gorai D, Mondal A, Sarkar S, Majhi S. A new labdane diterpene from *Rauvolfia tetraphylla* Linn. (Apocynaceae). Journal of Chemical Research 2011; 35(12): 678-680.

- [82]. Panda SK, Das D, Tripathy BN, Nayak L. Phytopharmacognostical studies and quantitative determination of reserpine in different parts of *Rauwolfia* (spp.) of Eastern Odisha by UV spectroscopy method. Asian Journal of Plant Science and Research 2012; 2(2): 151-162.
- [83]. Alagesaboopathi, C., "An investigation on the antibacterial activity of *Rauwolfia tetraphylla* dry fruit extracts" Ethanobotanical Leaflets, 2009,13: 644-650.
- [84]. Abubacker, M.N. and Vasantha, S. "Antibacterial activity of ethanolic leaf extract of *Rauwolfia tetraphylla* and its bioactive compound reserpine" Drug Invention Today, 2011, 3(3): 16-17.
- [85]. Shariff, N., Sudarshana, M.S. et al "Antimicrobial activity of *Rauwolfia tetraphylla* and *Physalis minima* leaf and callus extracts" Afr. J. Biotechnol, 2006, 5(10): 946-950.
- [86]. Suresh, K., Saravana, B. et al "Studies on in-vitro antimicrobial activity of ethanol extract of Rauwolfia tetraphylla" Ethnobotanical Leaflets, 2008, 12: 586-590
- [87]. Ganga, B.R., Umamaheswara, P.R. et al "Evaluation of *in-vitro* antibacterial activity and anti-inflammatory activity for different extracts of *Rauwolfia tetraphylla* L. Root bark" Asian Pac J Trop Biomed, 2012, 2(10): 818-821.
- [88]. Goutam, B., Mandal, L.C. et al "A new labdane diterpene from *Rauwolfia tetraphylla*" Journal of Chemical Research, 2011, 35(3): 678-680.
- [89]. Srivastava SK, Agrawal AK, Singh SC, Khanna VK, Singh J, Nath C, et al. IN 0658DEL2009, WO/2010/113180; 2010.
- [90]. Szewczak MR, Cornfeldt ML, Dunn RW, Wilker JC, Geyer HM, Glamkowski EJ, et al. Drug Dev Res 1987;11:157.
- [91]. Shikha, G., Vinayak, K.K. et al "Bioactive guided isolation of antipsychotic constituent from the leaves of *Rauwolfia tetraphylla* L" Fitoterapia, 2012, 83: 1092-1099.
- [92]. Vadakkemuriyil, D. N., Rajaram, P. and Ragupathi, G., "Studies on methanolic extract of *Rauwolfia* species from Southern Western Ghats of India- *In-vitro* antioxidant properties, Characterisation of nutrients and phytochemicals" Industrial Crops and Products, 2012, 39: 17-25.
- [93]. Gangarao, B., Umamaheswara, R. et al., Evaluation of *in-vitro* antibacterial activity and anti-inflammatory activity for different extracts of *Rauvolfia tetraphylla* L. root bark, Asian Pac J Trop Biomed, 2012, 2(10): 818-821
- [94]. Anonymous. The wealth of India: A Dictionary of Indian Raw Materials and Industrial Products, Publications and Information Directorate. New Delhi: CSIR; 2003; 166.
- [95]. Parrota, J.A., Healing plants of Penisular India, New York, USA: CABI Publishers, 2001, 120.
- [96]. Villar, R., Calleja, J.M. et al Screening of 17 Guatemalan medicinal plants for platelet antiaggregant activity, Phytother Res, 1997, 11: 441-445.