Cassia Fistula Seeds Nutritional Profile; An Insight into its Therapeutic Potentials

Oyewole T. A. Department of Science Technology Biochemistry unit, Federal Polytechnic,Ado-Ekiti, Ekiti State Nigeria Oyewole O.N. Department of Science Technology Chemistry unit, Federal Polytechnic, Ado- Ekiti, Ekiti State, Nigeria Falodun A.E. Central Research Laboratory, Federal Polytechnic, Ado Ekiti, Ekiti State, Nigeria

Abstract:- Cassia fistula has long been thought as ornamental plant due to its attractive leaves and the seeds has been regarded as waste product particularly in Nigeria. However, due to its numerous therapeutic attributes, it has remarkably gained popularity in Asian and African countries. Therefore, this study aimed to assess the important biological metabolites inherent in the Cassia fistula seeds using high throughput analytical instrumentations. The investigated seeds appeared to be good sources of magnesium, potassium, calcium, phosphorus, zinc iron, but the selenium and copper deficient when compared content were with **Recommended Dietary Allowance. The essential amino** acids contents were found to be relatively high in the seeds with highest values of Arginine of 8.90605%. The essential amino acid profiles of seeds were compared favorably with FAO/WHO (1991) requirement pattern when compared to recommended amino acids scores. Fatty acid profiles revealed that the seed samples is rich in monounsaturated fatty acids with values of 31.8658% and polyunsaturated fatty acids (44.9563%) with low saturated fatty acids. The study provides evidence thatCassia fistulaseeds contain essential amino acids, minerals which could be of interest for the development of new drugs and it could be valuable as an additives, supplement for animal feeds, nutraceuticals and therapeutic medicine.

Keywords:- *Fatty acids, phytochemicals, amino acids, bioactive compounds.*

I. INTRODUCTION

Cassia fistula is a tree plant commonly found in Africa, Asia and some parts of the world. It is a tracheophytes plants of the family of Fabacea with a scientific name *Cassia fistula*(NPDC *et al.*, 2000), and is commonly calledGolden shower (Bhalodia*et al.*, 2012). Despite the traditional claims of the potency of the different parts of *Cassia fistula* in traditional medicine, more scientific research work could be done on its pharmacological activities in human and poultry animals. The description of the *Cassia fistula* plant parts are shown in Figure 1.The leaves, barks, pulps etc. of these plants have been used by traditional practitioners in the treatment of different diseases (Ploeger and Shugar, 2017).

It has also been shown that extracts prepared from certain plants could be used as pill, decoction and powder for the management of several diseases (Xiong, 2003). *Cassia fistula* seeds have been reported to be useful in the management of jaundice (Asolkaret al., 1992), skin disorders, gastric distress, swollen throat and many more among some tribes (Bodding, 1983). Evidences have shown that some bioactive compounds in this plant could play key roles in pharmacological activities; increasing lifespan of mice with tumor (Gupta *et al.*, 2000), lowering blood sugar thereby revealing its anti-diabetic property (Bhakta *et al.*, 1997), hepatoprotective (Bhakta *et al.*, 1999), antioxidant (Luximon-Ramma*et al.*, 2002) activities among many others. Also, it has been reported to possess anti-fungal activity due to its clearance profile as anti-candidal agents (Subramanion*et al.*, 2012).

Researchers have isolated, identified and investigated numerous chemical compounds, and their applications in the prevention and management of diseases have been established (Fabricant and Farnsworth, 2001).Some primary and secondary metabolites present in different seed plants have been documented to play vital Pharmacological activities on human health and even poultry foods.

One of such are the protein building blood called the amino-acids with diverse metabolic roles such as; in the synthesis of protein, they help to stimulate insulin secretion in pancreatic β -cells stimulating glucose clearance from the blood in the management and prevention of Diabetes mellitus. Other amino-acids like arginine, leucine, isoleucine, alanine and phenylalanine have been reported to possess this insulin stimulating effects (Chen *et al.*, 2016; Birech*et. al.*, 2017).

Moreover, polyunsaturated fatty acids are the unit of fats essential during pregnancy, early child growth and development, during lactation for the development of the brain at early life. Evidences have linked PUFA to play vital role in the management of coronary heart related diseases when SFA was replaced with PUFA reducing coronary heart diseases in Adult, older men and women (Jakobsen*et al.*, 2009). The anti-carcinogenic properties of some conjugated linolenic acid and monounsaturated fatty acids in animal models have also been reported (Ve´ronique*et al.*, 2008).

In addition, some crop plant seeds and their oil have been reported to be rich source of polyunsaturated fatty acids (PUFA) essential in the human diet and in the management of diseases such as cardiovascular related diseases, cancer related diseases, neurological and hormonal disorders (Viorica-Mirela*et al.*, 2012).

Several minerals from different plant food and parts such as Zn, Cu, Mg, Ca are essential for proper health functioning and since mineral deficiency has been a major health challenge in different part of the world, increasing dietary mineral is a great therapy in combating these challenges in human and even other an (Dureaj*et al.*, 2003).

Also, Plants, herbs containing vitamins, minerals, amino-acids used in traditional medicine have been used as

a remedy to manage diseases and deficiencies (Pandey *et al.*, 2011).

There is little information on the therapeutic and health related profiles of these C seeds in Nigeria. Therefore Gas chromatography method was used to determine the fatty acid profiles, amino acid profiles of these seeds, which will help to unfold the medicinal and other importance of these seeds in Nigeria and other parts of the World.



Fig. 1: Diagram of the different plant parts of *Cassia fistula* where; (A). Is the tree showing the branches and leaves (B). Are the pods (C). Are the pods revealing the internal pulps and seeds (D). Are the pulps (E). Are the seeds

II. MATERIALS AND METHODS

Plant Material *Cassia fistula* seed pods were collected from The Federal Polytechnic Ado-Ekiti, Ekiti- State Nigeria. The pods were allowed to dry and the seeds were removed from them and washed with water. The seeds were then air-dried for five days after which it was grinded into powder and stored in an airtight container and all chemicals used were of analytical grade.

A. Phytochemical analysis

The qualitative and quantitative analysis of Secondary metabolites of aqueous extracts of *Cassia fistula* seeds was carried out using standard methods as described by Sofowora (1993) and Harborne (1973).

B. Determination of mineral composition

Analysis of mineral element was determined according to the official method of the Association of Official Analytical Chemists (AOAC, 1990). The sample analysis digestion was carried out using aqua regia (nitric acid and hydrochloric acid mixture in ratio 1:3). 15 ml concentrated acid was added to 1 g of sample in a conical flask of 100 ml, and then make to boil by heating on a fumed cupboard until the solution becomes whitish. The filtered solution was made up to 50 ml with distilled water then transferred into a sample bottles and labeled. The digested sample mineral analysis was determined using atomic absorption spectrophotometer (AAS) (Buck Scientific AAS Model 211 VGP) and Jenway Digital Flame Photometer (PFPT Model).

C. Fatty acid analysis using Gas chromatography

C seed fatty acid analysis were carried out using GC-flame ionization detection (GC-FID) (HP 6890 Powered with HP ChemStation Rev. A 09.01[1206] Software, with a FID detector equipped with a 30 m \times 0.25 mm \times 0.25 µm HP INNO Wax column dimension. The temperatures of the injection port and the detector were 250 and 320°C, respectively. Samples were desorbed in the split mode (split ratio 20:1). The oven temperature program was initially held at 60°C and first rampling at 12°C/min for 20 minutes, maintained for 2 minutes and the second rampling at 15°C/minutes for 3 minutes was maintained for 8 minutes. Nitrogen was used as the carrier gas. The peak areas of the target compounds were used to quantify the absolute contents compared to that of calibration samples with known concentrations.

D. Amino acid analysis

Amino Acid profile Extraction and analysis were carried out following the modified method of AOAC method 982.30, 2006 and Danka *et al.*, (2012) in the sample. 0.5 g of sample was weighed into the 250ml conical flask capacity. Fat content of the sample was defatted by extracting it with 30ml of petroleum spirit three times with thimble equipped soxhlet extractor. For complete hydrolysis to be achieved, the sample was hydrolysed three times for the totality of amino acids recovery. 30ml of the 1M potassium hydroxide was used to soak the pulverized and defatted sample and solution was incubated for 48 hours at 110°C in hermetically closed borosilicate glass container. Hence, the hydolysate was neutralized to get pH in the range of 2.5- 5.0 after the Alkaline hydrolysis. Cation exchange solid-phase extraction was used to purify the solution. The amino acids in purified solution were derivatised with ethylchloroformate before injection to Gas chromatography.

III. RESULT

A. Qualitative phytochemical analysis

The qualitative analysis of *Cassia fistula* seed aqueous extracts carried out base on standard protocols, with alkaloid, tannin, saponin, flavonoid, steroid, terpenoid, phenol and anthraquinone are present in the plant while glycoside is absent (Table 1).

Secondary metabolites	Test	Inference
Alkaloid	Mayer's test	+ve
Tannin	Ferric chloride test	+ve
Glycoside	Keller's test	-ve
Saponin	Frothing test	++ve
Flavonoid	Ferric chloride test	+++ve
Steroid	Salkowski test	+ve
Terpenoid	Salkowski test +ve	
Phenol	Ferric chloride test	++ve
Anthraquinone	Carbon tetrachloride test	+ve
Table 1: Phytochemical analysis of C seed		

Absent

+ Trace

++ Moderate

+++ Abundant

B. Quantitative phytochemical analysis

The *Cassia fistula* seeds aqueous extract shows higher concentration of phenol, followed by flavonoid terpenoids and tannin (Table 2).

Secondary metabolites	Values (%)	
Alkaloid	1.0170	
Terpenoid	2.5000	
Tannin	1.2468	
Flavonoid	4.8581	
Phytate	0.2131	
Phenol	13.410	

Table 2: Phytochemical composition of *C* seed

C. Mineral analysis of Cassia fistula seed

The mineral analysis of the *Cassia fistula* seeds using the atomic absorption spectrophotometer (AAS). The level of potassium is higher followed by magnesium, phosphorus and calcium as shown in (Table 3).

SAMPLE	SEEDS (ppm)	
Sodium (Na)	9.19	
Calcium (ca)	134.36	
Potassium (K)	319.36	
Iron (Fe)	9.1011	
Manganese (Mn)	0.7540	
Zinc (Zn)	6.1113	
Phosphorus (P)	149.11	
Magnesium (Mg)	171.080	
Selenium (Se)	0.7315	
Copper (Cu)	0.003	

Table 3: Mineral analysis of digested sample of *C* seeds

D. Fatty acid methyl ester analysis

The result and chromatograms of the fatty acid methyl ester analysis of *Cassia fistula* seed aqueous extracts using GC-flame ionization detection (GC-FID). The level of saturated fatty acid is 23.2%, monounsaturated fatty acid level is 31.9% and polyunsaturated fatty acid level is 45% as shown in (Table 4.0).

NAME	Amount/Area	Norm(%)	Area (pA*s)	
SATURATED FATTY ACID (SFA) METHYL ESTERS OF C seed				
Caprylic Acid (C8:0)	0.00000	0.00000	52.01508	
Capric acid (C10:0)	0.00000	0.00000	32.42331	
Lauric Acid (C12:0)	0.00000	0.00000	44.99215	
Myristic Acid (C14:0)	0.00600	0.31509	46.03571	
Palmitic Acid (C16:0)	0.01022	20.6507	130.7257	
Arachidic Acid (C20:0)	0.00815	0.95938	103.1966	
Behenic Acid (C22:0)	0.00334	0.48405	127.0199	
Lignoceric Acid (C24:0)	0.00469	0.76866	39.01125	
TOTAL 23.1779				
MONOUNSATURA	TED FATTY ACID(MU	FA) METHYL ESTERS OF	C seed	
Palmitoleic Acid (C16:1)	0.00468	0.89369	167.2208	
Margaric Acid (C17:1)	0.00132	0.05468	133.7457	
Stearic Acid (C18:1)	0.00637	7.12911	266.2669	
Oleic Acid (C18:1)	0.02321	23.4372	240.5085	
Erucic Acid (C22:1)	0.00530	0.35109	58.04701	
TOTAL		31.8658		
POLYUNSATURATED FATTY ACID (PUFA) METHYL ESTERS OF C seed				
Linoleic Acid (C18:2)	0.01126	43.6959	251.2725	
Linolenic Acid (C18:3)	0.00460	1.17625	60.7953	
Arachidonic Acid (C20:4)	0.00092	0.08410	79.97420	
TOTAL		44.9563		

Table 4: The Quantity Of Different Classes Of Fatty Acid Methyl Esters Profiles In C Seed

E. Amino acid analysis

The result and chromatograms of the amino-acids analysis of *Cassia fistula* seed aqueous extracts using Gas chromatography (GC-FID) are shown in (Table 5.0, Table 5.1 and Figure 2).

AMINO ACID	AMOUNT(g/100g Protein)
VALINE	5.06210
THREONINE	3.56616
ISOLEUCINE	3.91792
LEUCINE	7.37122
LYSINE	6.61034
METHIONINE	1.03464
PHENYLALANINE	6.66896
HISTIDINE	2.68106
ARGININE	8.90605
TYPTOPHAN	1.41660

Table 5: shows the Essential amino-acids quantities of C seed

AMINO ACID	AMOUNT(g/100g Protein)
GLYCINE	3.01746
SERINE	4.42244
PROLINE	5.44611
ALANINE	5.97326
GLUTAMATE	18.9154
ASPARTATE	8.61434
TYROSINE	2.68313
CYSTINE	1.10342

Table 6: shows the Non-essential amino acids quantities of *Cassia fistula* seed

The seed of this plant could be an important tool in discovering therapeutic drugs through isolation of some of these essential metabolites inherent in the seed.

IV. DISCUSSION

The bioactive constituents of *Cassia fistula* seeds were analyzed and identification of phytochemicals, minerals, saturated, monounsaturated, polyunsaturated fatty acid, and amino acids content were determined using standard procedures (Table 1-5).

The phytochemical screening of the aqueous extracts of *Cassia fistula* seed revealed the presence of phenol, saponin, tannin (Table 1-2) some of which have been documented to possess antioxidant, anti-inflammatory and antimicrobial activity in some fruits, vegetables and herbs (Pandey, 2015; Zhang *et al.*, 2015), Flavonoid in some plants, has been recorded to show vasodilating action against cardiovascular diseases (Zhang *et al.*, 2015).

The saponin contents of some plants have been reported to show hypolipidemic activity, anti-microbial activity, antiatherosclerotics (Yu et al., 2007), and antimicrobial activity in different studies (Kundan and Anupam, 2011).

Calcium, phosphorus, magnesium, potassium etc. are present in larger amounts compare to other mineral constituents present in *Cassia fistula* seed in (Table 3). Manganese, zinc, copper and selenium are co-enzyme of antioxidant enzymatic systems involve in defense against free radicals. Reduced intakes of certain micronutrients have been reported to increase oxidative stress in the pathogenesis of several diseases (Diplock, 1991).

The Gas chromatography analysis of fatty acid methyl ester show high level of PUFA and MUFA in *Cassia fistula* seed unraveling its therapeutic potentials (Table 4).

Some evidences of PUFA-rich diets have been reported to reduce Total cholesterol and LDL-cholesterol but manage to increase HDL-cholesterol among subjects on solid food experiment (Clarke *et al.*, 1997). And since replacing saturated fat with MUFA in some designed trials, it has proved to improve the CVD markers profiles (Mozaffarian and Clarke, 2009), significantly reduced high blood pressure in overweight model and decrease Triglyceride level among subjects with Diabetes mellitus (Schwingshackl*et al.*, 2011.) The Gas chromatography analysis of Amino acid in *Cassia fistula* seeds revealed the presence of ten essential amino acids (Table 5.0) and eight non-essential amino acids (Table 5.1).

The use of Amino acids in the management and treatment of diverse diseases have been documented; Histidine has been reported to be used in the management of Arthritis, enhance blood flow and proper functioning of the nervous system (Marta *et al.*, 2003).

Reports have shown that Arginine provides the substrate for the synthesis of nitric oxide, a mediating factor of vascular homeostasis (Flammer*et al.*, 2012). Impairment of NO production or bioactivity has been considered in cardiometabolic risk, including that of coronary artery disease, stroke, and diabetes (Reriani*et al.*, 2010). Omega-3 fatty acids have been documented in the treatment of osteoarthritis and athreosclereosis (Miggiano and Gagliardi, 2005).

Plants due to their phytoconstituents have been a major key factor in the management of human diseases and improving human health over time. Traditional practitioners have always used these medicinal plants in the cure and treatment of diverse infections since old times. Due to these observations of the traditional applications of these plants in health related conditions, carrying out scientific investigations on some of the important bioactive constituents present in *Cassia fistula* seed was considerable bearing in mind the therapeutic values present in some different seed plants.

Several reports have shown the therapeutic values of different parts of *Cassia fistula* and such therapeutic and pharmacological values includes: anti-inflammatory, anti-microbial, anti-ulcer, anti-tusive, wound healing, antipyretic, antiparasitic, anti-itching activities (Maya *et al.*, 2012) among many others. Different metabolites inherent in different plant parts have been documented to be a major factor in these diverse activities in some part of the World.

Amino acids, minerals, vitamins, unsaturated fats etc. play vital roles against oxidative stress, supporting the growth and development of poultry animals and reducing their risk of diseases, regulating immune response and maintaining normal metabolic and physiological systems. Since there is a high demands for growth enhancement additives, disease preventing drugs etc. Checking the

amounts of these constituents in *Cassia fistula* seeds could be a step in the right direction (Alagawanya*et al.*, 2021)

Some of these constituents unravel the therapeutic potentials of the seeds as they have been reported to have immense benefits in the management of different diseases. The high level of some amino-acids and minerals have given information on the biological roles the seeds could play in disease treatment and in further isolation and synthesis of drugs. The seeds have shown characteristic possible potential and applications in Nutraceuticals and in drug design.

V. CONCLUSION

This plant is commonly use in other part of the World, however the importance of these tree plant have not maximized in Nigeria. The *Cassia fistula* seed is so rich in essential metabolites such as palmitic acid, linolenic acid, oleic acid, phenols, flavonoids, Arginine, phenylalanine, Valine, lysine, isoleucine and minerals such as K, Ca, Mg, P which have all proved to play different roles in the management of diseases are also present in these cassia fistula seeds.More information of the medicinal and nutritional values of the seed would and the detection of more bioactive compounds and isolating them could be a major step towards understanding the therapeutic actions of this seed in disease management.

REFERENCES

- [1.] Asolkar, L.V., Kakkar, K.K., Chakre, O.J., (1992). Glossary of Indian Medicinal Plants with Active Principles, vol. 1. CSIR, New Delhi, pp. 177–178
- [2.] Bhakta, T.; Mukherjee, P.K.; Mukherjee, K.; Banerjee, S.; Mandal, S.C.; Maity, T.K.; Pal, M.; Saha, B.P (1999). Evaluation of hepatoprotective activity of Cassia fistula leaf extracts. J. Ethnopharmacol. 66, 277–282.
- [3.] Bhakta, T.; Mukherjee, P.K.; Saha, K.; Pal, M.; Saha, B.P (1997). Hypoglycemic activity of C Linn. (Leguminosae) leaf (Methanol extract) in alloxan-induced diabetic rats. J. Ethnobot. 9, 35–38.
- [4.] Birech Z, Mwangi PW, Bukachi F, Mandela KM (2017). Application of Raman spectroscopy in type 2 diabetes screening in blood using leucine and isoleucine amino-acids as biomarkers and in comparative anti-diabetic drugs efficacy studies. PLoS ONE 12(9): e0185130.
- [5.] Chen T, Ni Y, Ma X, Bao Y, Liu J, Huang F, Hu C, Xie G, Zhao A, Jia W (2016). Branched-chain and aromatic amino-acid profiles and diabetes risk in Chinese populations Scientific Reports 6: 20594.
- [6.] Clarke, R.; Frost, C.; Collins, R.; Appleby, P.; Peto, R (1997). Dietary lipids and blood cholesterol: Quantitative meta-analysis of metabolic ward studies. BMJ 316, 112–117.
- [7.] Danka petrovaObreshkova, DobrinaDonchevaTsvetkova and kallinValentinovIvanov (2012). Simultaneous indentification and determination of total content of Amino acid in food supplement- Tablets by Gas

chromatography. Asian journal of pharmaceutical and clinical Research, Vol 5, Sppl 2, 2012.

- [8.] Diplock A.T (1991). Antioxidant nutrients and disease prevention: an overview. Am J Clin Nutr.;53 (1 Suppl):189S-193S.
- [9.] Dureaj H, Kaushik D and Kumar V (2003), Development of nutraceuticals, Indian Journal of Pharmacology, 2003; 35: 363-72.
- [10.] Fabricant, D. S. and Farnsworth, N. R. (2001). The value of plants used in traditional medicine for drug discovery. Environ. Health Perspect. 109 (Suppl 1): 69–75.
- [11.] Flammer AJ, Anderson T, Celermajer DS, Creager MA, Deanfield J, Ganz P, Hamburg NM, Luscher TF, Shechter M, Taddei S, (2012). The assessment of endothelial function: from research into clinical practice.Circulation 2012;126(6):753–67.
- [12.] Gupta, M.; Mazumder, U.K.; Rath, N.; Mukhopadhyay, D.K (2000). Antitumor activity of Methanolic extract of C L. seed against Ehrlich Ascites Carcinoma. J. Ethnopharmacol. 2000, 72, 151–156.
- [13.] Harborne, J.B (1973): Phytochemical Methods: A Guide to Modern Techniques of Plant Analysis. Chapman A & Hall.London. Pp 279.
- [14.] Jakobsen, M. U., O'Reilly, E. J., Heitmann, B. L., Pereira, M. A., Balter, K., Fraser, G. E., et al. (2009).
- [15.] Major types of dietary fat and risk of coronary heart disease: A pooled analysis of 11 cohort studies. The American Journal of Clinical Nutrition, 89, 1425– 1432.
- [16.] Luximon-Ramma, A.; Bahorun, T.; Soobrattee, M.A.; Aruoma, O.I. Antioxidant activities of phenolic, proanthocyanidin, and flavonoid components in extracts of Cassis fistula. J. Agric. Food Chem. 2002, 50, 5042–5047.
- [17.] M Gupta; U.K Mazumder; N Rath; D.K Mukhopadhyay (2000). Antitumor activity of methanolic extract of C L. seed against Ehrlich Ascites Carcinoma. Journal of Ethnopharmacology, 72(1-2), 0–156.
- [18.] Mahmoud Alagawanya ,Shaaban S. Elnesrb , Mayada R. Faragc , RuchiTiwarid , Mohd. Iqbal Yatooe ,Kumaragurubaran Karthikf , Izabela Michalakg and Kuldeep Dhamah (2021). A Comprehensive Review on Nutritional significance of amino acids, vitamins and minerals as nutraceuticals in poultry production and health. Veterinary Quarterly, Vol, 41 No, 1, 1-29
- [19.] Marion Burckhardt, Max Herke, Tobias Wustmann, Stefan Watzke, Gero Langer, Astrid Fink, 2016.Omega-3-fatty acid for the treatment of dementia. Cochcrane Database of systemic reviews 4(4):CD009002
- [20.] Marta AH, Luigi F, Nathalie M, Floriana S, Angelo T (2003). Designing the Selectivity of the Fluorescent Detection of Amino Acids: A Chemo sensing Ensemble for Histidine. Jour. Am. Chem. Soc;125(1): 20–21
- [21.] Maya Kushwaha, Ramesh Chandra Agrawal (2012).Biological activities of the plant Cassia. Journal of Scientific Research in Pharmacy, Review Article ISSN: 2277-9469.

- [22.] Miggiano GA, Gagliardi L (2005). Diet, nutrition and rheumatoid arthritis. ClinTer 2005;156:115–23
- [23.] Mozaffarian, D.; Clarke, R (2009). Quantitative effects on cardiovascular risk factors and coronary heart disease risk of replacing partially hydrogenated vegetable oils with other fats and oils. Eur. J. Clin. Nutr. 63, 22–33.
- [24.] N. R. Bhalodia, R. N. Acharya, and V. J. Shukla (2012), "Evaluation of in vitro antioxidant activity of hydroalcoholic seed extrates of C Linn," Free Radicals and Antioxidants, vol. 1, no. 1, pp. 68–76,
- [25.] National Plant Data Center (NPDC), NRCS, USDA (2000). Baton Rouge, LA 70874-4490 USA. <u>http://plants.usda.gov</u>
- [26.] Nisreen Husain, TouseefHussainTrak and LataMeshram (2019). Amino acids as Medical food and their Therapeutic uses. *International Journals of Scientific Research and Reviews*, 8(2), 579-585
- [27.] Pandey N, Prasad M R, Rai S K, Rai S P (2011). Medicinal plants derived nutraceuticals: A reemerging health aid. Int J Pharm Bio Sci., 2011; 2: 420-41.
- [28.] Pandey S. Preliminary phytochemical screening and in vitro antibacterial activity of Bauhinia variegata Linn. against human pathogens. Asian Pac J Trop Dis 2015; 5: 123-9.
- [29.] R. Ploeger, A. Shugar, The story of Indian yellow excreting a solution, J. Cult. Herit. 24 (2017) 197–205.
- [30.] Reriani M.K, Lerman LO, Lerman A (2010). Endothelial function as a functional expression of cardiovascular risk factors. BiomarkMed ;4(3):351–60.
- [31.] Schwingshackl, L.; Strasser, B.; Hoffmann, G (2011). Effects of monounsaturated fatty acids on cardiovascular risk factors: A systematic review and meta-analysis. Ann. Nutr. Metab. 2011, 59, 176–186.
- [32.] Sofowora A (1993): Medicinal Plants and Traditional Medicines in Africa. Chichester John Willey & Sons New York. Pp.256.
- [33.] Subramanion L. Jothy, ZurainiZakariah, Yeng Chen, and SreenivasanSasidharan (2012). In Vitro, in Situ and in Vivo Studies on the Anticandidal Activity of *C* Seed Extract. Molecules 2012, 17, 6997-7009.
- [34.] Ve'roniqueChaje`s, Anne C. M. Thie'baut, MaximeRotival, Estelle Gauthier, VirginieMaillard, Marie-Christine Boutron-Ruault, VirginieJoulin, Gilbert M. Lenoir, and FrancxoiseClavel-Chapelon (2008). Association between Serum trans-Monounsaturated Fatty Acids and Breast Cancer Risk in the E3N-EPIC Study. American Journal of Epidemiology Vol. 167, No. 11.
- [35.] Viorica-MirelaPopa, Alexandra Gruia, Diana-nicoleta Raba, Delia Dumbrava, Camelia Moldovan, DespinaBordean, ConstantinMateescu (2012). Fatty acids composition and oil characteristics of linseed (LinumUsitatissimum L.) from Romania. Journal of Agroalimentary Processes and Technologies, 18 (2), 136-140.
- [36.] Xiong S. (2003). Medicinal-use extract of Medicago sativa root. Faming Zhuanli Shenqing Gongkai Shuomingshu, 6.

- [37.] Yu-Jie Zhang , Ren-You Gan , Sha Li, Yue Zhou, An-Na Li, Dong-Ping Xu and Hua-Bin Li (2015). Review on Antioxidant Phytochemicals for the Prevention and Treatment of Chronic Diseases. *Molecules*, 20, 21138– 21156.
- [38.] Kundan Singh Bora and Anupam Sharma, (2011). A review on Phytochemical and pharmacological potential of *Medicago sativa*. Pharmaceutical Biology, 2011; 49(2): 211–220
- [39.] Yu C, Guo J, Shen H, Liang D, Zhao N, Wen J. (2007). Method for preparing saponin extract of Medicago sativa and its application in manufacturing hypolipidemic agents and antiatherosclerotics. Faming Zhuanli Shenqing Gongkai Shuomingshu, 1